

<https://helda.helsinki.fi>

Review : Alternative and novel feeds for ruminants: nutritive value, product quality and environmental aspects

Halmemies-Beauchet-Filleau, A.

2018-12

Halmemies-Beauchet-Filleau , A , Rinne , M , Lamminen , M , Mapato , C , Ampapon , T , Wanapat , M & Vanhatalo , A 2018 , ' Review : Alternative and novel feeds for ruminants: nutritive value, product quality and environmental aspects ' , Animal , vol. 12 , no. Supplement 2 , pp. S295-S309 . <https://doi.org/10.1017/S1751731118002252>

<http://hdl.handle.net/10138/309590>

<https://doi.org/10.1017/S1751731118002252>

cc_by_nc_nd

acceptedVersion

Downloaded from Helda, University of Helsinki institutional repository.

This is an electronic reprint of the original article.

This reprint may differ from the original in pagination and typographic detail.

Please cite the original version.

Review: Alternative and novel feeds for ruminants - nutritive value, product quality and environmental aspects

A. Halmemies-Beauchet-Filleau¹, M. Rinne², M. Lamminen^{1,3}, C. Mapato⁴, T. Ampapon⁴, M. Wanapat⁴ and A. Vanhatalo^{1,3}

¹*Department of Agricultural Sciences, FI-00014 University of Helsinki, Finland*

²*Production Systems, Natural Resources Institute Finland (Luke), FI-31600 Jokioinen, Finland*

³*Helsinki Institute of Sustainability Science, FI-00014 University of Helsinki, Finland*

⁴*Tropical Feed Resources Research and Development Center, Department of Animal Science, Faculty of Agriculture, Khon Kaen University, Khon Kaen 40002, Thailand*

Corresponding author: anni.halmemies@helsinki.fi

Short title: Alternative and novel feeds for ruminants

Abstract

Ruminant-based food production faces currently multiple challenges such as environmental emissions, climate change and accelerating food-feed-fuel competition for arable land. Therefore, more sustainable feed production is needed together with the exploitation of novel resources. In addition to numerous food industry (milling, sugar, starch, alcohol or plant oil) side streams already in use, new ones such as vegetable and fruit residues are explored, but their conservation is challenging and production often seasonal. In the temperate zones, lipid-rich camelina (*Camelina sativa*) expeller as an example of oilseed by-products has potential to enrich ruminant

26 milk and meat fat with bioactive *trans*-11 18:1 and *cis*-9,*trans*-11 18:2 fatty acids and
27 mitigate methane emissions. Regardless of the lower methionine content of alternative
28 grain legume protein relative to soybean meal (*Glycine max*), the lactation performance
29 or the growth of ruminants fed faba beans (*Vicia faba*), peas (*Pisum sativum*) and
30 lupins (*Lupinus* sp.) are comparable. Wood is the most abundant carbohydrate
31 worldwide, but agroforestry approaches in ruminant nutrition are not common in the
32 temperate areas. Untreated wood is poorly utilised by ruminants because of linkages
33 between cellulose and lignin, but the utilisability can be improved by various processing
34 methods. In the tropics, the leaves of fodder trees and shrubs (e.g. cassava (*Manihot*
35 *esculenta*), *Leucaena* sp., *Flemingia* sp.) are good protein supplements for ruminants.
36 A food-feed production system integrates the leaves and the by-products of on-farm
37 food production to grass production in ruminant feeding. It can improve animal
38 performance sustainably at smallholder farms. For larger-scale animal production,
39 detoxified jatropha (*Jatropha* sp.) meal is a noteworthy alternative protein source.
40 Globally, the advantages of single-cell protein (bacteria, yeast, fungi, microalgae) and
41 aquatic biomass (seaweed, duckweed) over land crops are the independence of
42 production from arable land and weather. The chemical composition of these feeds
43 varies widely depending on the species and growth conditions. Microalgae have shown
44 good potential both as lipid (e.g. *Schizochytrium* sp.) and protein supplements (e.g.
45 *Spirulina platensis*) for ruminants. To conclude, various novel or underexploited feeds
46 have potential to replace or supplement the traditional crops in ruminant rations. In the
47 short-term, N-fixing grain legumes, oilseeds such as camelina and increased use of
48 food and/or fuel industry by-products have the greatest potential to replace or
49 supplement the traditional crops especially in the temperate zones. In the long-term,

50 microalgae and duckweed of high yield potential as well as wood industry by-products
51 may become economically competitive feed options worldwide.

52

53 **Keywords:** legume, by-product, single-cell protein, tree, ruminant

54

55 **Implications**

56 Within ruminant-based food production, there are potential means to improve global
57 food supply and to decrease its environmental footprint without compromising animal
58 products. Alternative and novel feeds provide opportunities to (a) spare arable land,
59 fresh water (e.g. single-cell proteins, duckweed) or fertilizers (N-fixing grain and shrub
60 legumes), (b) exploit side streams more efficiently (residues of food, biofuel or wood
61 production) and (c) increase the use of fibrous feeds not suitable for monogastrics
62 (wood, shrubs). They may also offer additional benefits such as modification of lipids
63 in ruminant products (lupins, camelina, microalgae) and mitigation of methane
64 emissions (lipid-rich feeds, tropical shrubs).

65

66 **Introduction**

67 Ruminant-based food production faces currently multiple and global challenges such
68 as needs to respond to the growing human population and food security, but also to
69 the pollution of environment and the accelerating climate change. The animal
70 production sector is also heavily criticised due to food-feed competition *i.e.* the feeding
71 of human-edible materials to animals and the use of arable land to produce animal
72 feed instead of producing human-edible food directly. Recently increasing interest in
73 biofuel production tightens up the competition on the use of arable land.

74

75 Ruminants are often criticised for the lower feed conversion efficiency relative to
76 monogastric livestock, but taking into account differences in the feed rations modifies
77 the ranking order. Indeed, to produce the same amount of animal protein products
78 (meat, milk or eggs) much less human-edible feed is needed in ruminant systems than
79 in monogastric systems (6 vs. 16 kg of human-edible feed DM per kg of protein
80 products, Mottet *et al.*, 2017). The strengths inherent to ruminant animals in food
81 production chain could be further developed by more diverse and efficient exploitation
82 of side streams and increased exploitation of fibrous feeds not suitable for the nutrition
83 of humans and monogastric livestock. To improve the food system sustainability and
84 to reach climate change targets, changes in feed and animal production alone are not
85 adequate. Changes in food consumption as regard to wastage and balanced dietary
86 choices are also needed (Röös *et al.*, 2017). According to Schader *et al.* (2015),
87 feeding animals solely based on food industry by-products and grasslands combined
88 with changes in human dietary patterns (reductions of animal products) have potential
89 to decrease the environmental load of food production drastically. For example,
90 greenhouse gas emissions, nitrogen (N) and phosphorus (P) load, as well as land and
91 fresh water use could decrease up to 18-46%.

92

93 Almost half of worldwide bovine milk production takes place in the temperate areas of
94 Europe and Northern America (FAOSTAT, 2016) under intensive (high inputs including
95 concentrate, high milk yield) or extensive production systems (high forage, low inputs,
96 moderate or low milk yield). At the present, the ruminant milk and meat production in
97 Europe relies largely on imported soybean (*Glycine max*) from South America
98 (Lindberg *et al.*, 2016). Soybean together with cereals and maize (*Zea mays*), lucerne
99 (*Medicago sativa*) or grass forage are typical dietary ingredients in the intensive

farming of the temperate zones. However, the highest cattle populations are in the tropical and subtropical climate zones, the number of cattle in Brazil and India alone comprising 15 and 13% of global cattle population, respectively (FAOSTAT, 2016). In the tropics, the forages are typically of poor nutritive value in terms of low protein and high fibre content that limits the efficiency of animal production. Local protein sources are thus sought both in the temperate as well as tropical areas.

Enteric methane emissions from ruminants significantly contribute to the environmental footprint of agriculture (Herrero *et al.*, 2016). Ruminal methane production also represents a substantial loss of feed energy. Appropriate forage supplementation and feed choices to improve forage and total diet digestibility have significantly more potential to increase ruminant performance and mitigate methane emissions in the extensive than in the intensive ruminant production systems (Knapp *et al.*, 2014; Herrero *et al.*, 2016). Modern intensive agriculture is a significant source of N emissions as well. Globally, about 50% of the N fertilizer applied to conventional cropping systems is not utilised by plants, but lost to the environment as ammonia (NH₃), nitrate (NO₃⁻), and nitrous oxide (N₂O, Coskun *et al.*, 2017). Legumes with biological N₂ fixation (Watson *et al.*, 2017) may offer an environmentally sound and sustainable nutrient source to ruminants. Furthermore, the N use efficiency of ruminants is mainly determined by diet N content (Huhtanen *et al.*, 2008) indicating the potential to reduce N leakages by dietary N optimisation.

The feasibility of using alternative feeds for ruminants depends among others on the feed value of novel feeds, animal production responses and feed costs compared to the conventional feeds. In addition, the environmental footprint of feed and animal

production, and the economic value of novel feeds in alternative uses such as energy production are of great importance. The objective of this article is to review the nutritive value of some currently underutilised or novel feeds for ruminants in the temperate zones (intensive and extensive farming) and in the tropics (extensive farming). In addition, the effects of these feeds on ruminant milk production and quality (milk, protein and fat yields and milk fatty acid composition) as well as meat production (average daily gains and meat composition) are examined and compared to more conventional feeds. The environmental load of novel feeds is evaluated based on requirements for arable land and for fresh water during the feed production and their possible effects on methane and nitrogen emissions of ruminants. This review comprises a quantitative evaluation of replacing traditional feeds by alternative ones on ruminant milk production as well as a comparative estimation of time delay for novel feeds to enter readily on the market together with their future potential to increase sustainable production and utilisation in ruminant nutrition.

Intensive and extensive ruminant production in the temperate zones – protein and energy supplements

By-products of food and bioenergy industries

Numerous food and biofuel industry side streams are already used as major components of ruminant diets such as hulls and feed meals from milling industry, distillery and brewery by-products, meals and expellers from plant oil production, molasses and pulps from sugar processing etc. (Feedipedia, 2018; Luke, 2018). Biofuel by-products as ruminant feeds have been reviewed in detail by Makkar *et al.* (2012). Recent attempts have aimed at utilising such side streams that have not

previously been used. Wadhwa and Bakshi (2013) estimated that nearly 50% of all fruits and vegetables in the European Union go to waste with losses occurring during agricultural production, processing, distribution and by consumers. Vegetable residues may be composted and used as soil amendments but with only a limited added value. One option to add value to these products is to preserve them by sun drying (Wadhwa *et al.*, 2015) or ensiling (Orosz and Davies, 2015) and feed to livestock. Vegetable and fruit residues are challenging raw materials for ensiling as they are easily perishable and typically moist (Wadhwa *et al.*, 2015; Table 1; Supplemental Table S1). Solid-state fermentation of the fruit and vegetable wastes in combination with other non-competing human food biomass could possibly (a) enrich them with proteins and other nutrients, (b) improve feed quality and (c) enhance ensilability (Wadhwa *et al.*, 2015).

The production of fruit and vegetable residues is often seasonal, and in many cases they are produced by small or medium size companies, resulting in rather small batches. To be able to recycle these residues back into the food chain requires high hygienic quality of the products and good stability to allow efficient logistics. Some of the major constraints in the use of fruit wastes are the presence of antinutritional factors such as pesticides, mycotoxins, heavy metals and dioxins (Wadhwa *et al.*, 2015). There are however positive experiences as e.g. ensiled tomato and olive by-products have been successfully used in the diets of dairy goats (Arco-Pérez *et al.* 2017) and ensiled apple pomace up to 30% in the diets of lactating dairy cows (Wadhwa *et al.*, 2015).

[Please, add Table 1 near here]

By-products of oilseed crops such as soybean and rapeseed meals and expellers are widely used as supplementary protein for dairy cows. One of the less used oilseed crops is an ancient plant camelina (*Camelina sativa*). Camelina has a moderate seed yield potential (Table 2) that combined with low nutrient requirements and a good resistance to diseases, pests and drought makes it adapted also to low-input farming (Heuzé *et al.*, 2017b). Camelinaseed oil is an economically interesting on-farm raw material for biofuel production (Keske *et al.*, 2013) to increase farmers' energy independence. Camelinaseed oil is also fit for human consumption (Heuzé *et al.*, 2017b). Camelina expeller contains lipids with significant amounts of essential fatty acids 18:2n-6 and 18:3n-3 (Bayat *et al.*, 2015), but it is also relatively abundant in CP and essential amino acids (**AA**) (Table 1). However, ruminal degradability of camelina protein *in situ* (76%) was higher than that of soybean (58%) or rapeseed (52%; Lawrence and Anderson, 2015). Feeding unprocessed or processed camelinaseeds to ruminants has sometimes, but not always, decreased DM intake (Table 3; Supplemental Table S2; Table 4; Supplemental Table S3) that may be related to glucosinolates (Lawrence *et al.*, 2016). Nevertheless, replacing various conventional protein feeds in ruminant diets with camelina expeller has resulted in comparable milk and protein yields (Table 3) or average daily gains (ADG, Table 4).

[Please, add Tables 2, 3 and 4 near here]

Feeding camelina expeller results in high concentrations of *trans*-11 18:1 and *cis*-9,*trans*-11 18:2, unaltered or slightly decreased 18:0 and *cis*-9 18:1 concentrations and a significant decrease in total saturated fatty acids in dairy cow (Halmemies-Beauchet-Filleau *et al.*, 2011 and 2017), in sheep (Szumacher-Strabel *et al.*, 2011) and in goat milk (Cais-Sokolińska *et al.*, 2015) as well as in sheep meat (Table 4). Besides

beneficially modifying lipids in ruminant milk and meat, camelina lipids at inclusion rate of 6% in the diet DM decreased ruminal methane and carbon dioxide production of dairy cows by 29 and 34%, respectively (Bayat *et al.*, 2015). However, caution should be exercised in the dosage of lipids as the reduction in methane emissions due to the dietary polyunsaturates may be accompanied with lowered DM intake and milk yield (Bayat *et al.*, 2015).

Grain legume seeds

Grain legumes such as faba bean (*Vicia faba*), pea (*Pisum sativum*) and lupins (*Lupinus* sp.) are old crops cultivated in all arable continents. There are three major modern lupine species bred to animal feed namely white (*Lupinus albus*), blue (*Lupinus angustifolius*) and yellow lupin (*Lupinus luteus*). In the short-term, grain legumes are presumably the most promising alternatives to soybean (*Glycine max*) and rapeseed in the temperate areas because their cultivation practices are already available and implemented (Figure 1). However, grain legume seeds are edible by humans as well. Therefore, the utilisation of human-inedible feeds for ruminants and/or feeds the production of which require less or not at all arable land should be encouraged to improve further the sustainability of food production system in the longer term.

The unique capacity of leguminous plants in conjunction with rhizobium symbionts to biologically fix and utilise atmospheric N enables that inorganic N-fertilisers with rising prices and high requirement of energy in manufacturing are not required. Indeed, the emissions of a potent greenhouse gas N₂O from legume cultivation are generally lower than those from N-fertilized crops (1.3 kg/ha vs. 3.2 kg/ha; Watson *et al.*, 2017). The

seed yield potential of grain legumes under optimal conditions is similar or exceeding that of conventional protein crops (Table 2). These advantages make legumes increasingly attractive in the intensive farming in addition to current wide spread use in the low-input and organic farming.

A prerequisite for the spread of grain legume production is the profitability relative to other crops. This is influenced e.g. by yields, volatile producer prices, incentives and production costs. Though the producer prices of grain legume seeds are on average 1.1 to 2.0 times higher than that of wheat in Europe (FAOSTAT, 2016), the competitiveness against more common crops such as wheat is uncertain mainly due to inconsistent DM yields and high seed costs. However, the incentives for protein feeds and reducing the seed costs by producing the seed on-farm can improve the competitiveness of grain legume cultivation. The cultivation of grain legumes is more challenging than that of cereals and grasses as they are sensitive to lodging and due to pests and pathogens they require efficient crop rotation (van Krimpen *et al.*, 2013). Nevertheless, the plant breeding may be able to overcome these agronomical constraints if given enough attention and resources.

Grain legume seeds differ in the chemical composition, the CP content ranging from 240 (peas) to 400 g/kg DM (soybeans). Soybeans have in general the highest ether extract (**EE**) content, whereas faba beans and peas contain significant amounts of starch and lupin seeds NDF (Table 1). The main storage carbohydrate of lupins is pectin instead of starch (White *et al.*, 2007). Lupin seeds contain more EE than faba beans and peas (Table 1) with *cis*-9 18:1 and 18:2n-6 as major fatty acids (White *et al.*, 2007). The protein in grain legume seeds, faba beans and lupin seeds in particular,

is low in methionine (Table 1), which is often the limiting AA for the lactation performance of dairy cows (e.g. Pisulewski *et al.*, 1996).

The feasibility of the use of alternative grain legumes in ruminant diets is determined not only by their chemical composition, but also by the rate and extent of degradation of nutrients in the rumen. The degradability of faba bean, pea and lupin protein in the rumen is often over 80% (Watson *et al.*, 2017) that is significantly higher than those of soybean or rapeseed expellers. In addition, the heat-treatment of faba beans, peas or lupin seeds to lower ruminal degradability has seldom improved animal performance (White *et al.*, 2007; Watson *et al.*, 2017). It is plausible that the high protein degradability in the rumen together with suboptimal AA profile in the undegraded protein of alternative grain legume seeds limit their production responses in high-yielding ruminants. Faba beans contain also antinutritional factors such as vicine and convicine (Heuzé *et al.*, 2016a), lupins quinolizidine alkaloids (Wasilewko and Buraczewska, 1999) and peas lectins and tannins (Heuzé *et al.*, 2017a). However, ruminants are not susceptible to most of them because of microbial metabolism and degradation in the rumen (Watson *et al.*, 2017).

Replacing protein in soybean meal partially or completely with faba beans, blue lupin, white lupin or peas has resulted in rather similar bovine lactation performances (Watson *et al.*, 2017; Table 3). Furthermore, the milk fat concentration of medium chain saturates has been lower and those of *cis*-9 18:1 and 18:2n-6 higher in cows fed white lupins seeds relative to soybean meal (White *et al.*, 2007). In contrast, the milk production responses of alternative grain legumes are often inferior compared to the rapeseed meal in dairy cow nutrition (Watson *et al.*, 2017; Table 3). Substitution of

rapeseed meal with faba beans has typically decreased milk protein yield and increased milk urea concentration and the proportion of N excreted in urine suggesting less efficient use of protein in faba beans than in rapeseed (Puhakka *et al.*, 2016; Table 3), thus leading to increased N emissions from animals.

Partial or total replacement of soybean or rapeseed protein by faba beans, lupin seeds or peas has not significantly altered ADG or meat chemical composition in growing sheep or cattle (Table 4). Besides replacing protein in ruminant diets, starchy faba beans and peas (Table 1) and lupins with higher metabolizable energy content than cereals (Watson *et al.*, 2017) have potential in replacing cereals as well. Indeed, the substitution of cereal grains by grain legumes in dairy cow diets generally increases milk production (White *et al.*, 2007; Watson *et al.*, 2017). Furthermore, starch in peas and faba beans has lower degradability in the rumen than cereal starch (Watson *et al.*, 2017) that lowers the risk for acidosis.

Biorefining of forage crops

Intrest in using grass biomass as a raw material for green biorefineries has arisen recently (McEniry and O'Kiely, 2014; Hermansen *et al.*, 2017). Grass is effective in converting solar radiation into chemical forms of energy and it grows well in humid temperate areas with a capacity for higher biomass and CP production compared to most annual crops (Table 2). Further, existing technology is available for its cultivation, harvesting and ensiling (Wilkinson and Rinne, 2018). When preserved as silage, the grass biomass can be refined all year round although losses in the protein and water soluble carbohydrates will take place during the fermentation process compared to the parent herbage.

299

300 Typically the first step in a green biorefinery process is liquid-solid separation resulting
301 in a liquid fraction containing the soluble components of grass and a fibrous solid
302 fraction. The yield of the fractions depends on the technical solutions of the process,
303 but it is also greatly affected by the raw material characteristics. The ensiling process
304 can even serve as a pretreatment for the biorefinery process, and it may be further
305 improved by using fibrolytic enzymes at the time of harvest as it has increased the
306 liquid yield (Rinne *et al.*, 2017). In the simplest approach, grass juice can be used as
307 a liquid feed to enrich the diet with highly nutritive forage based component and it is
308 readily consumed by dairy cows and monogastric animals (Rinne *et al.*, 2018), or the
309 fibre fraction can be used as a feed for ruminants (Savonen *et al.*, 2018). Grass fibre
310 is less lignified than e.g. woods and straw, and milder processes can be used to
311 hydrolyse it (Niemi *et al.*, 2017). The hydrolysed sugars can further be used for a
312 variety of purposes including direct use as feeds, and as substrates for lactic acid
313 fermentation or single-cell protein production. Green biorefineries have potential to
314 improve local nutrient self-sufficiency, provide new business opportunities for rural
315 communities and to produce ecosystem services such as improved soil structure,
316 carbon sequestration and biodiversity. The high costs related to transportation and
317 processing have to date prevented the development of commercial green biorefineries
318 on a large scale (Xiu and Shahbazi, 2015).

319

320 **Intensive and extensive ruminant production in the temperate zones – fibrous**
321 **feeds**

322

323 *Grain legumes as forage*

Harvesting grain legume stands as whole crop silage enables the utilisation of nutrients in stems and leaves as well and extending the cultivation in areas where the length of growing season may limit complete seed ripening. Although yield potential and organic matter digestibility of grain legume stands are high (Rinne *et al.*, 2014; Table 2), data on the effects of grain legume whole crop silages on ruminant performance and product quality is limited. In milk production, white lupin silage resulted in lower total DM intakes, but almost similar bovine lactation performance to maize silage as basal forage (Kochapakdee *et al.*, 2002). In meat production, animal performance has been similar or better when white lupin or pea silages have replaced partially or completely grass silage in cattle or sheep diets (Table 4). Due to their lower fibre concentration relative to grass silage, legume silages may lower ruminal methane emissions (Hristov *et al.*, 2013).

Compared to sole cropping, the bi-cropping of grain legumes and cereals may enhance and stabilize DM yields, reduce weeds and plant diseases and improve N-fixation (Hauggaard-Nielsen *et al.*, 2008). As a forage, grain legume-cereal crop mixtures complement the nutritive value of each other providing an appropriate balance between readily fermentable nutrients and N in the rumen (Watson *et al.*, 2017). Replacing half of the grass silage DM with faba bean-wheat silage had no effect on DM intake or bovine milk, fat and protein yields or feed N conversion efficiency to milk protein (Lamminen *et al.*, 2015). Whole crop faba bean-wheat or pea-wheat silages have successfully replaced grass silage in beef production as well (Table 4). Due to the lower costs of N fertilizers and good yield potential, grain legume silages seem to provide a viable alternative for maize and grass silages both in the intensive and extensive production systems (Table 2). The feeding value and ruminal methane

emissions of diets containing forage legumes (lucerne, clovers) have been reviewed elsewhere (Dewhurst, 2013).

Temperate wood-derived products

Wood is the most abundant source of carbohydrates worldwide. Principal components of wood are cellulose (400 to 450 g/kg DM) and hemicelluloses (200 to 300 g/kg DM, Sjöström, 1993). Agroforestry approaches in ruminant nutrition are less common in the temperate areas compared to the tropics or the Mediterranean area. There are however some applications where e.g. willow (*Salix* sp.) production for wood chips and the grazing of ruminants are combined to provide additional benefits such as improved microclimate for the animals, self-medication and soil carbon sequestration, although the potential of the untreated wood based materials to provide energy and nutrients to high yielding dairy cows is limited (Smith *et al.*, 2012, 2014). Indeed, the *in vitro* digestibility of DM of untreated wood of various tree species was poor with a range from 0.002 to 0.035 (Millett *et al.*, 1970).

A variety of technologies have been used over decades to improve the digestibility of wood derived lingo-cellulosic materials. The key is to break the link between the lignin and the cell wall carbohydrates, particularly hemicelluloses, in order to improve the digestibility of ligno-cellulose by rumen microbes. Most pulping and papermaking residues have undergone at least partial delignification. Depending on the process, the residue may contain different proportions of hemicellulose and/or cellulose with or without lignin. The digestibility of pure cellulose is rather high and corresponds to the digestibility of typical ruminant feeds such as cereal grains and good quality forages. Saarinen *et al.* (1959) determined the *in vivo* digestibility of 40 wood pulps produced

by various pulping methods and reported a range in digestibility from 0.27 to 0.90 depending on the lignin content. The *in vivo* digestibility of bleached (lignin erased and the pulp whitened) chemical pulp fines from mixed hardwood was 0.78 for DM and 0.86 for carbohydrates (Millett *et al.*, 1973), indicating that the materials have a high energy value for ruminants.

Although wood derived cellulose can be used as a feed for ruminants, it has higher value as e.g. paper raw material. In contrast, hemicelluloses are a by-product of pulping that are typically burned, and interest of using them as feeds has arisen. Hemicelluloses are not homogeneous compounds but a group of mixed polysaccharides. They can be divided into four groups according to their main type of sugars: xylans, xyloglucans, mannans and β -glucans. Spruce (*Picea* sp.) and pine (*Pinus* sp.; softwood) contain somewhat less hemicelluloses than birch (*Betula* sp.; hardwood) and hemicellulose composition differs between species (Saarinen *et al.*, 1959). Glucomannans and galactomannans are the principal hemicelluloses of coniferous trees (spruce and pine) and xylans in deciduous trees (birch) while β -glucans are restricted to grasses.

Hemicelluloses in a liquid form are often called wood molasses or wood sugar concentrates. They have successfully been used as diet components for ruminants at up to 10% of DM intake (Zinn *et al.*, 1990 and 1993; Herrick *et al.*, 2012). An *in vitro* gas production experiment revealed that hot water and pressure extracted galactoglucomannan and xylan were readily used as fermentation substrates by rumen microbes of dairy cows fed a grass silage and cereal based diet but arabinogalactan was not (Rinne *et al.*, 2016). In an *in vivo* digestibility trial, the organic matter

digestibility (**OMD**) of the hot water and pressure extracted galactoglucomannan was 0.591 (Rinne *et al.*, 2016).

Bark is another component of wood that has limited value in the pulp and sawmill industry. Although wild ruminants consume bark voluntarily, the energy value of it is so low that incorporating it into dairy cow diets resulted in the reduction of milk production (P. Kairenius *et al.*, unpublished results). Thus, some processing would be needed to improve the digestibility of bark. Wood derived feeds typically have very low N and P concentrations. If the basal diet were high in these nutrients, wood derived feeds could dilute diets and subsequently increase e.g. the N use efficiency of lactating dairy cows as it is mainly determined by N intake (Huhtanen *et al.*, 2008). Wood derived feeds may also provide a source of feed in the case of lack of other feeds e.g. in crisis situations. In general, they may fit best in the diets of animals with low energy requirements rather than in dairy cow diets in the intensive production systems.

Extensive ruminant production in the tropics – protein supplements

Fodder trees and shrubs

Low quality forages such as rice (*Oryza sativa*) straw and pangola (*Digitaria eriantha*) grass low in protein and high in NDF and ADF are common in ruminant nutrition in the tropics (42, 691 and 424 g/kg DM for rice straw (Heuze and Tran, 2015b) and 5-12, 610-790 and 350-420 g/kg DM for pangola grass (Tikam *et al.*, 2013), respectively). Thus, the basal diet is typically much lower in protein and higher in fibre compared to that used in the intensive ruminant production of the temperate zones. In Asian tropics, rice straw is commonly supplemented with cassava (*Manihot esculenta*) chip rich in

soluble carbohydrates but poor in CP (750 to 850 g/kg DM and 20 to 30 g/kg DM, respectively; Wanapat and Kang, 2015) and soybean meal. However, the high price of soybean meal limits its use in smallholder farming.

Leaves of local fodder trees and shrubs such as cassava, leucaena (*Leucaena leucocephala*), moringa (*Moringa oleifera*) and sesbania (*Sesbania sesban*) often contain almost as much CP as NDF (Table 1), the concentration of former being roughly half of that in soybean meal. Supplementing the rice straw based diets with these alternative protein sources increases DM intake, improves microbial protein synthesis in the rumen and the efficiency of rumen fermentation with a shift towards propionate (Table 5; Supplemental Table S4), thus potentially mitigating methane production. These beneficial changes may be due to certain natural secondary compounds present in these alternative feeds, namely condensed tannins and saponins (Wanapat *et al.*, 2013).

[Please, add Table 5 near here]

Combined food-feed production system to provide a year round feeding calendar and to enrich smallholder farming environment is illustrated in Supplemental Figure S1. Under the proposed system, two grass types with (a) erect and tall growth habit and (b) semi-prostrate or prostrate growth habit are used to maximise the biomass production under zero-grazing and grazing, respectively. Roots from cassava can be utilised as a carbohydrate source while the whole top is dried to provide protein (Wanapat, 2009; Wanapat *et al.*, 2017). Additionally, the leaves of fodder trees and shrubs such as leguminous leucaena, flemingia (*Flemingia macrophylla*), and moringa

are harvested in intervals and used fresh or preserved for later use. The intercropping of cassava with leguminous crops, e.g. common bean (*Phaseolus calcaratus*) and cowpea (*Vigna unguiculata*), has potential to improve soil fertility and to increase biomass yield (Wanapat, 2009; Wanapat *et al.*, 2017). Crop residues such as rice straw, corn stover and sugar cane top are also exploited in ruminant feeding.

Jatrophas

Jatrophas are drought-resistant shrubs or small trees native to American tropics and widely distributed in the tropical and subtropical regions around the world. *Jatropha* genus includes more than 175 species, *J. curcas* being one of the most studied species in animal feeding. *Jatropha* is an interesting biofuel crop due to the high EE concentration of its kernels (570-600 g/kg DM; Makkar *et al.*, 2012), and the de-fatted kernel residue, *jatropha* kernel meal, is a good source of nutrients with CP concentration of 620 to 770 g/kg DM (Table 1). In comparison to soybean protein, *jatropha* is deficient in lysine, but richer in other essential AA (Table 1; Makkar *et al.*, 2012).

The majority of *jatropha* species are highly toxic to both ruminants and monogastrics due to phorbol esters (1-3 mg/g kernel meal; Makkar *et al.*, 2012), but they can successfully be detoxified. The complete detoxification is absolutely necessary to avoid animal mortality (Elangovan *et al.*, 2013). In addition, the high concentration of antinutritional factors (trypsin inhibitors, lectin and phytate) may limit the use of *jatropha* especially for monogastrics unless deactivated by heat treatment and supplemented with phytase enzyme. When completely detoxified, the substitution of

soybean by jatropha has not impaired the DM intake or ADG of sheep and goats (Table 4). Though the yield potential is high (Table 2), the inconsistency of yields of current cultivars is the major restriction for the spread (Heuzé *et al.*, 2016b).

All production systems of ruminants worldwide – alternative protein and fibrous feeds

The major advantages of single-cell protein, seaweed and duckweed are the independence of production from arable land and of weather conditions as well as the high and continuous harvests (Nasseri *et al.*, 2011; van der Spiegel *et al.*, 2013; Table 2). However, cultivation, harvesting, preservation (especially drying) and application in feed in a large scale needs further research (van Krimpen *et al.*, 2013) to lower the production cost of these novel feeds to competitive level. In the long-term, microalgae and duckweed have perhaps the greatest potential to become viable local protein and fibre sources for ruminants worldwide (Table 2; Figure 1).

Single-cell protein

Single-cell protein consists of microbial cells from yeast, bacteria, fungi or microalgae. These micro-organisms can utilise a wide variety of inexpensive feedstocks and wastes as sources of carbon, nutrients and energy for growth to produce biomass rich in protein. The protein content of SCP varies due to culture conditions, species and strains (Lindberg *et al.*, 2016) but is in the same order as in soybean expeller (Table 1). The major constraints are the risk for allergens and the accumulation of heavy metals, pesticides and toxins especially if grown on polluted and contaminated substrates, generally high nucleic acid content (bacteria and yeasts > fungi >

microalgae; 60-120, 70-100, 30-80 g/kg DM, respectively) and economical and efficient mass-scale production and harvesting (Nasseri *et al.*, 2011; Lindberg *et al.*, 2016). Dietary nucleic acids and their derivatives are rapidly degraded in the rumen and certain end-products can be re-used as sources of carbon and N for bacterial growth (McAllan, 1982), but the N in nucleic acids is not as easily available as that of true protein or ammonia.

The basic stages of SCP production process include (a) medium preparation, (b) fermentation or photosynthesis and (c) harvesting and downstream processing like washing, cell disruption, protein extraction and purification (Ravindra, 2000). The SCP concept was introduced already during the First World War primarily as a human food (Lindberg *et al.*, 2016). However, the higher production costs of SCP linked to challenges in efficient and economical cell recovery in relation to more conventional foods and feeds is perhaps the main reason why SCP has not reached widespread commercial use so far. Established processes include the use of yeasts *Candida lipolytica* and *C. tropicalis* with alkanes as substrate (product called Toprina), bacterium *Methylophilus methylophilus* with methane as substrate, bacterium *Pseudomonas methylotrophus* (Pruteen) with methanol as substrate, filamentous fungus *Peecilomyces variotii* grown on sulphite spent liquor of forest industry sidestream (Pekilo) and yeast *Kluveromyces marxianus* grown on whey (Nasseri *et al.*, 2011). The reasons why the SCP concept could become more common and economically viable in future are the rising ecoawareness and the need to intensify nutrient and resource utilisation combined with the sharp price rises caused by the prospect of protein scarcity (Lindberg *et al.*, 2016).

Microalgae

Microalgae are a diverse group of unicellular or simple multicellular microorganisms with widely varying nutritive composition (Table 1). As animal feed, microalgae have several potential uses. Species high in lipids, such as 22:6n-3-enriched *Schizochytrium* sp., can be used to modify ovine (Bichi *et al.*, 2013) or bovine (Boeckaert *et al.*, 2008) milk fat healthier for humans in terms of increased *trans*-11 18:1, *cis*-9,*trans*-11 18:2 and n-3 content. Algal 22:6n-3 supplementation has increased also the n-3 content of ruminant meat (Meale *et al.*, 2014), but no effects were found on methane production (Moate *et al.*, 2013). In turn, microalgae or defatted microalgae residues high in CP (e.g. *Spirulina platensis* and *Chlorella vulgaris*), or high in carbohydrates can substitute conventional protein (Lamminen *et al.*, 2017) or energy feeds (van Emon *et al.*, 2015), respectively.

The AA composition of microalgae generally compares favourably to soybean meal (Becker, 2013) and rapeseed meal (Feedipedia, 2018; Luke, 2018), but may vary significantly between species (Table 1). However, in comparison to rapeseed meal and soybean meal, microalgae protein is often lower in histidine, which is typically the first AA limiting milk production on grass silage and cereal based diets (e.g. Vanhatalo *et al.*, 1999). The protein degradability of many microalgae species is suggested to be higher than that of rapeseed (Costa *et al.*, 2016; Lamminen *et al.*, 2017), soybean and cottonseed meals (Costa *et al.*, 2016), but this can possibly be affected by the growing and harvesting conditions of microalgae (Lodge-Ivey *et al.*, 2014). Compared to the conventional protein or energy feeds, large doses of microalgae or defatted microalgae residue may impact negatively on feed intake of ruminants depending on microalgae composition (van Emon *et al.*, 2015; Costa *et al.*, 2016; Lamminen *et al.*, 2016, 2017).

The palatability of microalgae can possibly be improved by feed processing, e.g. pelleting (Hintz *et al.*, 1966). Compared to rapeseed meal, microalgae have not affected milk yield, but decreased the milk protein yield of dairy cows in late lactation, which together with decreasing N utilisation for milk production suggests that the protein value of microalgae is possibly slightly lower than that of rapeseed meal (Lamminen *et al.*, 2017), but similar to soybean protein (Table 3).

The local on-farm production of microalgae in ponds or in closed photoreactors connected to animal drinking water system could lower the energy inputs of feed drying, preservation and transportation making microalgae cultivation in future a viable concept also in the extensive farming. Indeed, microalgae have successively been distributed through drinking water (Panjaitan *et al.*, 2010) to growing cattle grazing low quality grasses to improve microbial protein production in the rumen and diet digestibility (Panjaitan *et al.*, 2015). In addition, microalgal derived renewable biofuels have high potential to replace fossil fuels of diminishing reserves in future. The cost for the biofuels production from microalgae is not yet competitive with fossil fuels, but with advancing technologies and possible government incentives it may soon become profitable (Milano *et al.*, 2016) thus providing defatted microalgae residues for livestock in a mass-scale.

Seaweeds

Seaweeds are complex multicellular organisms growing in salt water or a littoral zone of marine environment (van der Spiegel *et al.*, 2013). They can be of many different shapes, sizes, colours and composition. Fresh seaweed contains very large amounts of water (700–900 g/kg DM) and needs to be consumed quickly or preserved by e.g.

drying or ensiling. Brown algae (*Phaeophyceae*) are of lesser nutritional value than red (*Rhodophyceae*) and green algae (*Chlorophyceae*) due to lower CP content (up to 140 vs. up to 500 and 300 g/kg DM, respectively). The protein content of marine seaweeds varies between seasons, but *in situ* rumen degradable protein remains unaffected with high inherent variability between algal species (24 to 51% of CP; Tayyab *et al.*, 2016). Protein in all seaweeds is typically deficient in essential AA except for methionine (Makkar *et al.*, 2016; Table 1).

Seaweeds are low in cellulose (about 40 g/kg DM) but rich in specific complex carbohydrates (e.g. alginate, laminarin and fucoidan). Step-wise increase in the levels of seaweeds in the diet may enable rumen microbes to adapt and utilise these compounds (Makkar *et al.*, 2016). Seaweeds concentrate heavy metals and minerals from seawater and contain several times the ash content of land plants that limits their gross energy value and requires regular monitoring (van der Spiegel *et al.*, 2013; Makkar *et al.*, 2016).

Makkar *et al.* (2016) have recently reviewed in detail the nutritive value of seaweed indicating that some species have the potential to contribute to the protein and energy needs of ruminants (e.g. *Macrocystis pyrifera*, *Palmaria palmata*, *Laminaria digitata*, *Ulva lactuca*), while others contain a number of bioactive compounds, which could be used as prebiotics for enhancing production and health status of animals (e.g. *Ascophyllum nodosum*). Moreover, some seaweed species have shown potential to mitigate ruminal methane production *in vitro* depending on the basal diet (Maia *et al.*, 2016). The seaweeds used for animal feeding can be cultivated or harvested in the wild (Table 4; Makkar *et al.*, 2016; Tayyab *et al.*, 2016) serving to mitigate nutrient

loading and to counteract eutrophication processes (Lindberg *et al.*, 2016). However, high collection rates in the wild have impaired the equilibrium of coastal ecosystems (Makkar *et al.*, 2016). In addition, increased cultivation of seaweeds may promote increased production of bromoform, a metabolic by-product of seaweeds that causes the depletion of atmospheric ozone layer (Carpenter and Liss, 2000).

Duckweeds

Duckweeds are monocotyledonous, small floating plants with no stems or true leaves of the botanical family *Lemnaceae* comprising of 4 genera (*Lemna*, *Spirodela*, *Wolffia* and *Wolffiella*). Duckweeds are found worldwide, but they grow best in stagnant water between 17.5 and 30°C (Heuzé and Tran, 2015a) and may have a 50% biomass increase every two days (van Krimpen *et al.*, 2013). Thus, duckweed is a potential novel nutrient source for herbivores worldwide. Only few studies have been performed on duckweed in ruminants (van der Spiegel *et al.*, 2013). Overall, duckweed is consumed well in both dried and fresh forms (Heuzé and Tran, 2015a) and it can supply a significant proportion of protein and other nutrients to animals with no significant adverse effects on performance (Cheng and Stomp, 2009; Zetina-Cordoba *et al.*, 2013).

The duckweed protein is much lower in essential AA histidine, methionine and lysine compared to that of soybean and rapeseed expeller (Table 1) that may limit duckweed's production responses relative to them. Estimates of ruminal protein degradability vary widely between 50 and 80% (Heuzé and Tran, 2015a). Duckweed contains significant amounts of ash and NDF (Table 1), but has low lignin content (57 g/kg DM; Heuzé and Tran, 2015a). It has therefore potential to substitute also forage

(Zetina-Cordoba *et al.*, 2013) and minerals (particularly P; van der Spiegel *et al.*, 2013) in ruminant diets. Nevertheless, high oxalic acid content may restrict the use of duckweed for livestock (van der Spiegel *et al.*, 2013).

Similarly to microalgae, local on-farm production of duckweed e.g. in ponds may offer a viable concept for ruminant feed production in future. Nutrient scavenging from field runoffs, manure and greywater by duckweeds has potential to reinforce circular economy practices at farm level and to decrease the environmental footprint of ruminant-based food production systems. The very high growth rate (van Krimpen *et al.*, 2013) enables that duckweed could be regularly harvested and fed to animals as fresh. Feeding fresh duckweed also limits the costs related to drying and preservation on-farm. Due to much bigger particle size relative to microalgae, simple mechanical harvesting of duckweed is feasible.

Conclusions

In the short term, the seeds and whole crop forages of N-fixing grain legumes as well as by-products from food and biofuel industries have the greatest potential to replace or supplement traditional crops in ruminant rations in the intensive and extensive production systems in the temperate zones (summarising Figure 1). Lipid-rich camelina expeller, as an example, beneficially modifies the fatty acid composition of ruminant products with potential to mitigate simultaneously enteric methane formation, whereas the oil fraction of seeds could be used as an on-farm biofuel to increase the energy independence of farmers. In the tropics, the leaves of fodder trees and shrubs (e.g. cassava, *Leucaena* sp., *Flemingia* sp.) are good protein supplements for ruminants especially in the extensive production systems where the potential to

improve diet digestibility and to mitigate enteric methane emissions is the highest. Combined food-feed production system to improve animal productivity and the efficiency of nutrient recycling as well as to decrease footprint on environment is recommended to smallholders (summarising Supplemental Figure 1), whereas detoxified jatropha meals could be suited for larger-scale feed and animal production in the tropics.

In the long-term, microalgae and duckweed of high yield potentials may become economically competitive local protein and fibre sources, respectively, for ruminants worldwide (Figure 1). This is due to the independence of their production from arable land and weather conditions while animal performance and product quality remain comparable to the traditional feeds. Microalgal derived renewable biofuels have a high potential to replace fossil fuels of diminishing reserves in future, thus providing defatted microalgae residues for intensive livestock farming in a mass-scale. Furthermore, on-farm production of microalgae connected to animal drinking water system could lower energy inputs of feed drying, preservation and transportation making microalgae competitive feed ingredient also in extensive farming. Exploitation of vast nutrient reserves in forests both in the temperate and tropical zones warrants further research on their feed value, the breaking of lignin-linkages of wood material and subsequent animal production responses.

Under the climatic conditions changing at an accelerating pace, the ruminant-based livestock systems in both temperate and tropical environments are very flexible in the types of biomasses that can be used as feeds. Despite the environmental footprint of ruminants, their importance in food production system cannot be ignored because of

their unique ability to naturally consume fibrous vegetable material not exploitable to humans and other monogastrics and convert it to milk and meat of high nutritive value. Transition to ruminant diets comprising fibrous feed sources supplemented exclusively on alternative and novel feeds has great potential to improve sustainability of ruminant-derived food production, which will not compete with human-edible food materials.

Acknowledgements

This review is presented in The International Symposium on the Nutrition of Herbivores (ISNH) 2018 in Clermont-Ferrand, France. The authors thank organising committee for the invitation.

References

- Arco-Pérez A, Ramos-Morales E, Yáñez-Ruiz DR, Abecia L and Martín-García AI 2017. Nutritive evaluation and milk quality of including of tomato or olive by-products silages with sunflower oil in the diet of dairy goats. *Animal Feed Science and Technology* 232, 57-70.
- Bayat AR, Kairenius P, Stefański T, Leskinen H, Comtet-Marre S, Forano E, Chaucheyras-Durand F and Shingfield KJ 2015. Effect of camelina oil or live yeasts (*Saccharomyces cerevisiae*) on ruminal methane production, rumen fermentation, and milk fatty acid composition in lactating cows fed grass silage diets. *Journal of Dairy Science* 98, 3166–3181.
- Becker EW 2013. Microalgae for human and animal nutrition. In *Handbook of microalgal culture: applied phycology and biotechnology* (ed. A. Richmond and Q Hu), second ed., pp. 461-503. Wiley-Blackwell, Chicester, United Kingdom.
- Bichi E, Hervás G, Toral PG, Lóor JJ and Frutos P 2013. Milk fat depression induced by dietary marine algae in dairy ewes: Persistency of milk fatty acid composition and animal performance responses. *Journal of Dairy Science* 96, 524-532.

697 Boeckaert C, Vlaeminck B, Dijkstra J, Issa-Zacharia A, Van Nespen T, Van Straalen W and
698 Fievez V 2008. Effect of dietary starch or micro algae supplementation on rumen fermentation
699 and milk fatty acid composition of dairy cows. *Journal of Dairy Science* 91, 4714-4727.

700 Cais-Sokolińska D, Pikul J, Wójtowski J, Danków R, Teichert J, Czyżak-Runowska G and
701 Bagnicka E 2015. Evaluation of quality of kefir from milk obtained from goats supplemented
702 with a diet rich in bioactive compounds. *Journal of the Science of Food and Agriculture* 95,
703 1343-1349.

704 Carpenter LJ and Liss PS 2000. On temperate sources of bromoform and other reactive
705 organic bromine gases. *Journal of Geophysical Research* 105, 20539-20547.

706 Cheng JJ and Stomp AM 2009. Growing duckweed to recover nutrients from wastewaters and
707 for production of fuel ethanol and animal feed. *Clean–Soil, Air, Water* 37, 17-26.

708 Coskun D, Britto DT, Shi W and Kronzucker HJ 2017. Nitrogen transformations in modern
709 agriculture and the role of biological nitrification inhibition. *Nature Plants* 3, 17074.

710 Costa DFA, Quigley SP, Isherwood P, McLennan SR and Poppi D 2016. Supplementation of
711 cattle fed tropical grasses with microalgae increases microbial protein production and
712 average daily gain. *Journal of Animal Science* 94, 2047-2058.

713 Dewhurst RJ 2013. Milk production from silage: comparison of grass, legume and maize
714 silages and their mixtures. *Agricultural and Food Science* 22, 57-69.

715 Elangovan AV, Gowda NKS, Satyanarayana ML, Suganthi RU, Rao SBN and Sridhar M 2013.
716 *Jatropha* (*Jatropha curcas*) seed cake as feed ingredient in the rations of sheep. *Animal*
717 *Nutrition and Feed Technology* 13, 57-67.

718 FAOSTAT, 2016. Retrieved on 22 April 2018, from <http://www.fao.org/faostat/en/#home>

719 Feedipedia 2018. Animal Feed Resources Information system. Retrieved on 27 April 2018,
720 from <http://www.feedipedia.org/>

721 Halmemies-Beauchet-Filleau A, Kokkonen T, Lampi AM, Toivonen V, Shingfield KJ and
722 Vanhatalo A 2011. Effect of plant oils and camelina expeller on milk fatty acid composition in
723 lactating cows fed diets based on red clover silage. *Journal of Dairy Science* 94, 4413–4430.

Halmemies-Beauchet-Filleau A, Shingfield KJ, Simpura I, Kokkonen T, Jaakkola S, Toivonen V and Vanhatalo A 2017. Effect of incremental amounts of camelina oil on milk fatty acid composition in lactating cows fed diets based on a mixture of grass and red clover silage and concentrates containing camelina expeller. *Journal of Dairy Science* 100, 305-324.

Hauggaard-Nielsen H, Jørnsgaard B, Kinane J and Jensen E 2008. Grain legume-cereal intercropping: the practical application of diversity, competition and facilitation in arable and organic cropping systems. *Renewable Agriculture and Food Systems* 23, 3-12.

Hermansen JE, Jørgensen U, Lærke PE, Manevski K, Boelt B, Jensen SK, Weisbjerg MR, Dalsgaard TK, Danielsen M, Asp T, Amby-Jensen M, Sørensen CA.G, Jensen MV, Gylling M, Leindedam J, Lübeck M and Fog E 2017. Green biomass – Protein production through bio-refining. DCA Report No. 093. Aarhus University, Denmark. 68 p. Retrieved on 25 November 2017, from www.dca.au.dk.

Herrero M, Henderson B, Havlík P, Thornton PK, Conant RT, Smith P, Wiersenius S, Hristov AN, Gerber P, Gill M, Butterbach-Bahl K, Valin H, Garnett T and Stehfest E 2016. Greenhouse gas mitigation potentials in the livestock sector. *Nature Climate Change* 6, 452-461.

Herrick KJ, Hippen AR, Kalscheur KF, Anderson JL, Ranatunga SD, Patton RS and Abdullah M 2012. Lactation performance and digestibility of forages and diets in dairy cows fed a hemicellulose extract. *Journal of Dairy Science* 95, 3342-3353.

Heuzé V and Tran G 2015a. Duckweed. Retrieved on 26 July 2017, from <https://www.feedipedia.org/node/15306>

Heuzé V and Tran G 2015b. Rice straw. Retrieved on 15 December 2017, from <https://feedipedia.org/node/557>

Heuzé V, Tran G, Delagarde R, Lessire M and Lebas F 2016a. Faba bean (*Vicia faba*). Retrieved on 26 July 2017, from <http://www.feedipedia.org/node/4926>

Heuzé V, Tran G, Edouard N, Renaudeau D, Bastianelli D, Lebas F 2016b. *Jatropha* (*Jatropha* sp.) kernel meal and other *jatropha* products. Retrieved on 23 April 2018, from <https://www.feedipedia.org/node/620>

752 Heuzé V, Tran G, Giger-Reverdin S, Noblet J, Renaudeau D, Lessire M and Lebas F 2017a.
 753 Pea seeds. Retrieved on 26 July 2017, from <http://www.feedipedia.org/node/264>
 754 Heuzé V, Tran G and Lebas F 2017b. Camelina (*Camelina sativa*) seeds and oil meal.
 755 Retrieved on 26 July 2017, from <http://www.feedipedia.org/node/4254>
 756 Hintz HF, Heitman H, Weir WC, Torell DT and Meyer JH 1966. Nutritive value of algae grown
 757 on sewage. Journal of Animal Science 25, 675-681.
 758 Hristov AN, Oh J, Firkins JL, Dijkstra J, Kebreab E, Waghorn G, Makkar HPS, Adesogan AT,
 759 Yang W, Lee C, Gerber PJ, Henderson B and Tricarico JM 2013. Special topics—Mitigation
 760 of methane and nitrous oxide emissions from animal operations: I. A review of enteric
 761 methane mitigation options. Journal of Animal Science 91, 5045-5069.
 762 Huhtanen P, Nousiainen JI, Rinne M, Kytölä K and Khalili H 2008. Utilization and partition of
 763 dietary nitrogen in dairy cows fed grass silage-based diets. Journal of Dairy Science 91,
 764 3589-3599.
 765 Keske CM, Hoag DL, Brandess A and Johnson JJ 2013. Is it economically feasible for farmers
 766 to grow their own fuel? A study of Camelina sativa produced in the western United States as
 767 an on-farm biofuel. Biomass and bioenergy 54, 89-99.
 768 Knapp JR, Laur GL, Vadas PA, Weiss WP and Tricarico JM 2014. Invited review: Enteric
 769 methane in dairy cattle production: Quantifying the opportunities and impact of reducing
 770 emissions. Journal of Dairy Science 97, 3231-3261.
 771 Kruus K and Hakala T 2016. The making of bioeconomy transformation. VTT Technicak
 772 Research Centre of Finland Ltd. Retrieved on 15 November 2017, from
 773 [https://makingoftomorrow.com/wp-content/uploads/2017/02/The-Making-of-Bioeconomy-](https://makingoftomorrow.com/wp-content/uploads/2017/02/The-Making-of-Bioeconomy-Transformation-2017.pdf)
 774 [Transformation-2017.pdf](https://makingoftomorrow.com/wp-content/uploads/2017/02/The-Making-of-Bioeconomy-Transformation-2017.pdf)
 775 Kochapakdee S, Moss BR, Lin J, Reeves DW, McElhenney WH, Mask P and Santen EV 2004.
 776 Evaluation of white lupin, temperate corn, tropical corn, and hybrid pearl millet silage for
 777 lactating cows. In Proceedings of the 10th International Lupin Conference, Wild and cultivated
 778 lupins from the Tropics to the Poles, 19-24 June 2002 Laugarvatn, Iceland, pp. 300-307.

779 Lamminen M, Kokkonen T, Halmemies-Beauchet-Filleau A, Termonen T, Vanhatalo A and
780 Jaakkola S 2015. Partial replacement of grass silage with faba bean whole-crop silage in the
781 diet of dairy cows. In Proceedings of the 18th Symposium of the European Grassland
782 Federation, Grassland and forages in high output dairy farming systems, 15-17 June 2015
783 Wageningen, The Netherlands, pp. 446-448.

784 Lamminen M, Halmemies-Beauchet-Filleau A, Kokkonen T, Jaakkola S and Vanhatalo A 2016.
785 Microalgae as a substitute for soya bean meal in the grass silage based dairy cow diets. In
786 Proceedings of 5th EAAP International Symposium on Energy and Protein Metabolism and
787 Nutrition, 12-15 September 2016, Krakow, Poland pp. 285-287.

788 Lamminen M, Halmemies-Beauchet-Filleau A, Kokkonen T, Simpura I, Jaakkola S, Vanhatalo
789 A 2017. Comparison of microalgae and rapeseed meal as supplementary protein in the grass
790 silage based nutrition of dairy cows. *Animal Feed Science and Technology* 234, 295-311.

791 Lawrence RL and Anderson JL 2015. Ruminal degradation and intestinal digestibility of
792 camelina and carinata meal compared with other protein sources. *Journal of Dairy Science*
793 98(Suppl. 2), 459.

794 Lawrence RD, Anderson JL and Clapper JA 2016. Evaluation of camelina meal as a feedstuff
795 for growing dairy heifers. *Journal of Dairy Science* 99, 6215-6228.

796 Lindberg JE, Lindberg G, Teräs J, Poulsen G, Solberg SØ, Tybirk K, Przedzimirska J, Sapota
797 GP, Olsen ML, Karlson H, Jóhannsson R, Smáráson BÖ, Gylling M, Knudsen MT, Dorca-
798 Preda T, Hermansen JE, Kruklite Z and Berzina I 2016. Nordic Alternative Protein Potentials:
799 Mapping of regional bioeconomy opportunities. Nordic Council of Ministers. Retrieved on 26
800 July 2017, from [http://www.nordic-ilibrary.org/environment/nordic-alternative-protein-](http://www.nordic-ilibrary.org/environment/nordic-alternative-protein-potentials_tn2016-527)
801 [potentials_tn2016-527](http://www.nordic-ilibrary.org/environment/nordic-alternative-protein-potentials_tn2016-527)

802 Lodge-Ivey SL, Tracey LN and Salazar A 2014. Ruminant nutrition symposium: The utility of
803 lipid extracted algae as a protein source in forage or starch-based ruminant diets. *Journal of*
804 *Animal Science* 92, 1331-1342.

805 Luke (Natural Resources Institute Finland) 2018. Feed tables and nutrient requirements.
806 Retrieved on 27 November 2018, from www.luke.fi/feedtables

807 Maia MRG, Fonseca AJM, Oliveira HM, Mendonça C and Cabrita ARJ 2016. The potential role
 808 of seaweeds in the natural manipulation of rumen fermentation and methane production.
 809 Scientific Reports 6, 32321.

810 Makkar HP, Cooper G, Weber JA, Lywood W and Pinkney J 2012. Biofuel co-products as
 811 livestock feed. Opportunities and challenges. Food and Agriculture Organization, Rome, Italy.

812 Makkar HP, Tran G, Heuzé V, Giger-Reverdin S, Lessire M, Lebas F and Ankers P 2016.
 813 Seaweeds for livestock diets: a review. Animal Feed Science and Technology 212, 1-17.

814 McAllan AB 1982. The fate of nucleic acids in ruminants. Proceedings of the Nutrition Society
 815 41, 309-316.

816 McEniry J and O’Kiely P 2014. Chapter 11: Developments in grass-/forage-based biorefineries.
 817 In Advances in biorefineries - Biomass and waste supply chain exploitation (ed. K Waldron)
 818 [Woodhead Publishing Series in Energy: Number 53][Library of Congress Control Number:
 819 2014931606; ISBN 978-0-85709-521-3 (print); ISBN 978-0-85709-738-5 (online)], pp. 335-
 820 363.

821 Milano J, Ong, HC, Masjuki HH, Chong WT, Lam MK, Loh PK and Vellayan V 2016. Microalgae
 822 biofuels as an alternative to fossil fuel for power generation. Renewable and Sustainable
 823 Energy Reviews 58, 180-197.

824 Millett MA, Baker AJ, Feist WC, Mellenberger RW and Satter LD 1970. Modifying wood to
 825 increase its in vitro digestibility. Journal of Animal Science 31, 781-788.

826 Millett MA, Baker AJ, Feist WC, Mellenberger RW and Satter LD 1973. Pulp and papermaking
 827 residues as feedstuffs for ruminants. Journal of Animal Science 37, 599-607.

828 Meale SJ, Chaves AV, He ML and McAllister TA. 2014. Dose–response of supplementing
 829 marine algae (*Schizochytrium* spp.) on production performance, fatty acid profiles, and wool
 830 parameters of growing lambs. Journal of Animal Science 92, 2202–2213.

831 Moate PJ, Williams RO, Hannah MC, Eckard RJ, Auldist MJ, Ribaux BE, Jacobs JL and Wales
 832 WJ 2013. Effects of feeding algal meal high in docosahexaenoic acid on feed intake, milk
 833 production, and methane emissions in dairy cows. Journal of Dairy Science 96, 3177–3188.

834 Mottet A, de Haan, C, Falcuccia A, Tempioa G, Opioa, C and Gerbera P 2017. Livestock: On
 835 our plates or eating at our table? A new analysis of the feed/food debate. *Global Food*
 836 *Security* 14,1-8.

837 Nasser AT, Rasoul-Amini S, Morowvat MH and Ghasemi Y 2011. Single cell protein:
 838 production and process. *American Journal of Food Technology* 6, 103-116.

839 Niemi P, Pihlajaniemi V, Rinne M and Siika-aho M 2017. Production of sugars from grass
 840 silage after steam explosion or soaking in aqueous ammonia. *Industrial Crops and Products*
 841 98, 93-99.

842 Orosz S and Davies DR 2015. Short and long term storage of wet by-products fed by
 843 ruminants. In *Proceedings of XVII International Silage Conference*, 1-3 July 2015, Piracicaba,
 844 Brazil. pp. 200-242.

845 Panjaitan T, Quigley SP, McLennan SR and Poppi DP 2010. Effect of the concentration of
 846 *Spirulina* (*Spirulina platensis*) algae in the drinking water on water intake by cattle and the
 847 proportion of algae bypassing the rumen. *Animal Production Science* 50, 405-409.

848 Panjaitan T, Quigley SP, McLennan SR, Swain AJ and Poppi DP 2015. *Spirulina* (*Spirulina*
 849 *platensis*) algae supplementation increases microbial protein production and feed intake and
 850 decreases retention time of digesta in the rumen of cattle. *Animal Production Science* 55,
 851 535-543.

852 Pisulewski PM, Rulquin H, Peyraud JL and Verite R 1996. Lactational and systemic responses
 853 of dairy cows to postruminal infusions of increasing amounts of methionine. *Journal of Dairy*
 854 *Science* 79, 1781-1791.

855 Puhakka L, Jaakkola S, Simpura I, Kokkonen T and Vanhatalo A 2016. Effects of replacing
 856 rapeseed meal with fava bean at 2 concentrate crude protein levels on feed intake, nutrient
 857 digestion, and milk production in cows fed grass silage-based diets. *Journal of Dairy Science*
 858 99, 7993-8006.

859 Ravindra P 2000. Value-added food: Single cell protein. *Biotechnology Advances* 18, 459-479.

860 Rinne M, Dragomir C, Kuoppala K, Smith J and Yáñez-Ruiz D 2014. Novel feeds for organic
 861 dairy chains. *Organic Agriculture* 4, 275-284.

- Rinne M, Kautto O, Kuoppala K, Ahvenjärvi S, Willför S, Kitunen V, Ilvesniemi H and Sormunen-Cristian R 2016. Digestion of wood-based hemicellulose extracts as screened by in vitro gas production method and verified in vivo using sheep. *Agricultural and Food Science* 25, 13-21. Retrieved on 15 December 2017, from <http://ojs.tsv.fi/index.php/AFS/article/view/46502>
- Rinne M, Winqvist E, Pihlajaniemi V, Niemi P, Seppälä A and Siika-aho M 2017. Fibrolytic enzyme treatment prior to ensiling increases press-juice yield from grass silage. In *Proceedings of the 8th Nordic Feed Science Conference*, 13-14 June 2017, Uppsala, Sweden. Swedish University of Agricultural Sciences, Department of Animal Nutrition and Management. Report 296. pp. 71-76. Retrieved on 15 December 2017, from <http://www.slu.se/globalassets/ew/org/inst/huv/nfsc/nfsc-2017-proceedings.pdf>
- Rinne M, Keto L, Siljander-Rasi H, Stefanski T and Winqvist E 2018. Grass silage for biorefinery – Palatability of silage juice for pigs and cows. Submitted to XVIII International Silage Conference, 24-26 July 2018, Bonn, Germany.
- Röös E, Bajželj B, Smith P, Patel M, Little D and Garnett T 2017. Protein futures for Western Europe: potential land use and climate impacts in 2050. *Regional Environmental Change*, 17, 367-377.
- Saarinen P, Jensen W and J Alhojärvi 1959. On the digestibility of high yield chemical pulp and its evaluation. *Acta Agralia Fennica* 94, 41-64.
- Savonen O, Franco M, Stefanski T, Mäntysaari P, Kuoppala K, Rinne M. 2018. Grass silage for biorefinery - Dairy cow responses to diets based on solid fraction of grass silage. *Nordic Feed Science Conference*, 12 -13 June 2018, Uppsala, Sweden.
- Schader C, Muller A, El-Hage Scialabba N, Hecht J, Isensee A, Erb K-H, Smith P, Makkar HPS, Klocke P, Leiber F, Schwegler P, Stolze M and Niggli U 2015. Impacts of feeding less food-competing feedstuffs to livestock on global food system sustainability. *Journal of Royal Society Interface* 12, 20150891.
- Sjöström E 1993. *Wood chemistry: fundamentals and applications*. Academic Press. USA. 293 p.

Smith J, Leach K, Rinne M, Kuoppala K and Padel S 2012. Integrating willow-based bioenergy and organic dairy production – the role of tree fodder for feed supplementation. In Proceedings of the 2nd IFOAM Animal Husbandry Conference, 12-14 September 2012, Hamburg, Germany. vTi Agriculture and Forestry Research, Special issue 362. pp. 417-420. Retrieved on 15 December 2017, from http://orgprints.org/21758/1/Smith_20AHC%20proceedings_2012.pdf

Smith J, Kuoppala K, Yáñez-Ruiz D, Leach K and Rinne M 2014. Nutritional and fermentation quality of ensiled willow from an integrated feed and bioenergy agroforestry system in UK. . In Proceedings of Maataloustieteen Päivät 2014, 8-9 January 2014, Helsinki, Finland. 9 p. Retrieved on 15 December 2017, from http://www.smts.fi/MTP_julkaisu_2014/Posterit/064Smith_ym_Nutritional_and_fermentation_quality_of_ensiled_willow.pdf

Szumacher-Strabel M, Cieślak A, Zmora P, Pers-Kamczyc E, Bielińska S, Stanisław M and Wójtowski J 2011. Camelina sativa cake improved unsaturated fatty acids in ewe's milk. Journal of the Science of Food and Agriculture 91, 2031-2037.

Tayyab U, Novoa-Garrido M, Roleda MY, Lind V and Weisbjerg MR 2016. Ruminant and intestinal protein degradability of various seaweed species measured in situ in dairy cows. Animal Feed Science and Technology 213, 44-54.

Tikam K, Phatsara C, Mikled C, Vearasilp T, Phunphiphat W, Chobtang J, Cherdthong A and Südekum KH 2013. Pangola grass as forage for ruminant animals: a review. SpringerPlus 2, 604-609.

USDA, 2016. Oil crops yearbook 2016. Retrieved on 15 November 2017, from <http://usda.mannlib.cornell.edu/MannUsda/homepage.do>

Van der Spiegel M, Noordam MY and Fels-Klerx HJ 2013. Safety of novel protein sources (insects, microalgae, seaweed, duckweed, and rapeseed) and legislative aspects for their application in food and feed production. Comprehensive Reviews in Food Science and Food Safety 12, 662-678.

917 Van Emon ML, Loy DD and Hansen SL 2015. Determining the preference, in vitro digestibility,
 918 in situ disappearance, and grower period performance of steers fed a novel algae meal
 919 derived from heterotrophic microalgae. *Journal of Animal Science* 93, 3121-3129.

920 Vanhatalo A, Huhtanen P, Toivonen V and Varvikko T 1999. Response of dairy cows fed grass
 921 silage diets to abomasal infusions of histidine alone or in combinations with methionine and
 922 lysine. *Journal of Dairy Science* 82, 2674–2685.

923 Van Krimpen MM, Bikker P, Van der Meer IM, Van der Peet-Schwering CMC and Vereijken
 924 JM 2013. Cultivation, processing and nutritional aspects for pigs and poultry of European
 925 protein sources as alternatives for imported soybean products (No. 662). Wageningen UR
 926 Livestock Research.

927 Wadhwa M and Bakshi MPS 2013. Utilization of fruit and vegetable wastes as livestock feed
 928 and as substrates for generation of other value-added products. *FAO Publication* 2013/04.
 929 H.P. Makkar Technical Editor. Retrieved on 15 December 2017, from
 930 <http://www.fao.org/docrep/018/i3273e/i3273e.pdf>.

931 Wadhwa M, Bakshi MP and Makkar HP 2015. Waste to worth: fruit wastes and by-products as
 932 animal feed. *CAB Reviews* 10, 1-26.

933 Wanapat M 2009. Potential uses of local feed resources for ruminants. *Tropical Animal Health*
 934 *and Production* 41, 1035–1049.

935 Wanapat M, Kang S and Polyorach S 2013. Development of feeding systems and strategies
 936 of supplementation to enhance rumen fermentation and ruminant production in the tropics.
 937 *Journal Animal Science and Biotechnology* 4, 32.

938 Wanapat M and Kang S 2015. Cassava chip (*Manihot esculenta* Crantz) as an energy source
 939 for ruminant feeding. *Animal Nutrition* 1, 266-270.

940 Wanapat M, Foiklang S, Ampapon T, Mapato C and Cherdthong T 2017. Feeding strategy on
 941 farms to improve livestock productivity and reduce methane production. In *Proceedings of*
 942 *the 2nd International Conference on Animal Nutrition and Environment*, 1-4 November 2017,
 943 Khon Kaen, Thailand, pp. 14-29.

944 Wasilewko J and Buraczewska L 1999. Chemical composition including content of amino
 945 acids, minerals and alkaloids in seeds of three lupin species cultivated in Poland. *Journal of*
 946 *Animal and Feed Sciences* 81, 1-12.

947 Watson CA, Reckling M, Preissel S, Bachinger J, Bergkvist G, Kuhlman T, Lindström K,
 948 Nemecek T, Topp CFE, Vanhatalo A, Zander P, Murphy-Bokern D and Stoddard F 2017.
 949 Chapter Four-Grain Legume Production and Use in European Agricultural Systems.
 950 *Advances in Agronomy* 144, 235-303.

951 White CL, Staines VE and Staines MvH 2007. A review of the nutritional value of lupins for
 952 dairy cows. *Australian Journal of Agricultural Research* 58, 185-202.

953 Wilkinson JM and Rinne M 2018. Review. Highlights of progress in silage conservation and
 954 future perspectives. *Grass and Forage Science*, 73: 40–52. doi 10.1111/gfs12327..

955 Xiu S and Shahbai A 2015. Development of green bio refinery for biomass utilization: A review.
 956 *Trends in Renewable Energy* 1, 4-15.

957 Zetina-Cordoba P, Ortega-Cerilla ME, Ortega-Jimenez E, Herrera-Haro JG, Sanchez-Torres-
 958 Esqueda MT, Reta-Mendiola JL, Vilaboa-Arroniz J and Munguia-Ameca G 2013. Effect of
 959 cutting interval of Taiwan grass (*Pennisetum purpureum*) and partial substitution with
 960 duckweed (*Lemna* sp. and *Spirodela* sp.) on intake, digestibility and ruminal fermentation of
 961 Pelibuey lambs. *Livestock Science* 157, 471-477.

962 Zinn RA 1990. Feeding value of wood sugar concentrate for feedlot cattle. *Journal of Animal*
 963 *Science* 68, 2598-2602.

964 Zinn RA 1993. Comparative feeding value of wood sugar concentrate and cane molasses for
 965 feedlot cattle. *Journal of Animal Science* 71, 2297-2302.

Table 1 Chemical composition of some alternative and common feeds for ruminants

Feed ¹	DM g/kg	Ash g/kg DM	NDF	Starch	EE ²	CP	His g/kg CP	Met	Lys
<i>Common protein feeds</i>									
Rapeseed expeller	899	69	299		92	391	28	22	56
Soybean expeller	907	68	111		77	493	27	14	63
<i>By-products of food industry</i>									
Apple pomace	360	26	525		50	77			
Camelinaseed expeller	905	69	305	2	156	357	23	20	46
Cauliflower leaf	654	162	145			126			
Cucumber waste	37	113	168			163			
Grape marc	876	63	658		64	115	29	15	45
Tomato fruit waste	62	101	191			103			
Olivesilage (pulp + leaf)	575	127	390			88			
<i>Grain legume seeds</i>									
Faba bean	866	39	159	447	14	290	26	8	62
Lupin, blue	915	42	253	122	63	332	28	7	50
Lupin, white	912	43	235	84	105	344	23	8	50
Lupin, yellow	898	54	254	35	53	435	27	7	50
Pea	865	35	142	513	12	239	25	10	72
Soybean	887	57	132	64	214	396	26	14	62
Grass silage juice	98	193				190			
<i>Grain legume whole crop stands</i>									
Faba bean	168	62	387	82		175			
Lupin, white	142	68	395			169			
Pea	198	65	397	67		167			
<i>Trees or shrubs (leaves unless otherwise stated)</i>									
Cassava	250	126	459			223		46 ³	
Flemingia	290	53	531			258		58 ³	
Leucaena	320	64	316			205		36 ³	
Moringa	330	115	219		54	251	31	21	66
Pine bark		22	667		47	28			
Sesbania	290	103	258			233			
Willow	264	71	573			167			
<i>Jatropha</i> kernel meal, detoxified	876-971	79-136	98-200	68-120	4-52	624-775	27-33	14-17	30-36
<i>Single-cell protein</i>									
Bacteria		30-70			10-30	500-650	23	30	61
Fungi		90-140			20-80	300-450	15-20	15-17	38-61
Microalgae									
<i>Chlorella vulgaris</i>	946	57	0	43	95	608	18	19	49
<i>Euglena gracilis</i>	960	35	0		138	240	26	20	66
<i>Scenedesmus obliquus</i>		60-100			120-140	500-600	15-17	12-21	50-57
<i>Schizochytrium</i> sp.		82	6 ⁴		380-710	121	8	< 8	33
<i>Spirulina platensis</i>	940	70	0	64	55	692	16	22	39
Yeast		50-100			20-60	450-550	21-22	13-21	74-77
<i>Seaweed</i>									
<i>Ascophyllum nodosum</i>	100-300	225	209		39	80	14	13	46
<i>Macrocystis pyrifera</i>	100-300	320	199		6	101	13	19	47
<i>Ulva</i> spp	100-300	230	262		12	186	20	16	38
<i>Duckweed</i>	56	159	401		61	291	17	8	39

¹ References in Supplemental Table S1 ² Ether extract ³ Tannins g/kg DM ⁴ Crude fibre

968 **Table 2** *The suitability for local production of some common and alternative feeds in different production systems, potential yields in*
969 *Europe, the need of land or water for feed production and other main environmental aspects regarding crop and ruminant production*

Feed	Local production ¹			Yield ² t/ha		Need for		Other environmental aspects
	TInt	TExt	Tropics	DM	N	Land	Fresh water	
Common feeds								
Rapeseeds	Yes	Yes		1.5-3	0.6-1.2	Arable	High	Need for N fertilization to get high yields. ²
Soybeans	Yes	Yes	Yes	3	0.8	Arable	High	Legume, but day length and temperature restricts yield potential and expansion to northern periphery. ²
Wheat	Yes	Yes	(Yes)	10	1.1	Arable	High	Need for N fertilization to get high yields.
Grass forage	Yes	Yes	(Yes)	10-15	1.2-2	Arable	High	Need for N fertilization to get high yields, or inclusion of forage legumes.
Alternative feeds								
Camelina seeds	Yes	Yes		3	0.8	Arable	High	Modest needs for cultivation compared to rapeseed. Polyunsaturates of Camelina lipid may decrease ruminal methane emissions. ³
Legume grains peas, beans, lupins	Yes	Yes	(Yes)	4-6	1-2	Arable	High	Legumes, therefore no need for N fertilization. High ruminal degradability of protein and unbalanced amino acid profile of undegradable protein may increase N emissions from ruminants. ⁴
Legume forage	Yes	Yes		13	2.5	Arable	High	Legumes, therefore no need for N fertilization. Due to lower fibre content, legume forages may mitigate ruminal methane emissions.
Hemicellulose		Yes	Yes			Forest	High	Low in N and P. Incorporation in the diet may improve N and P use efficiency if basal diet is excessive in these nutrients.
Leaves (tropical trees and shrubs)			Yes			Forest	High	Secondary compounds in certain species may direct rumen fermentation towards propionate and thus mitigate methane. ⁵
Jathropa fruit			Yes	2.5-5 ⁶	1.7-3.4 ⁶	Arable Forest	High	Decrease soil erodibility due to lateral roots. ⁶ Utilization of jathropa kernel meal that is a by-product of oil extraction as animal feed improves overall nutrient recycling.
Single-cell protein excluding microalgae	Yes	Yes	Yes			No	Low	Can recover nutrients from wastewaters and transform low-value organic by-products to feed.
Microalgae	Yes	Yes	Yes	15-30	4-15	No	Low	Can recover nutrients from wastewaters. Based on chemical composition, species rich in lipids and low in fibre may have

	Seaweed	(Yes)	(Yes)	25	2.5-7.5	No	No	potential to mitigate ruminal methane emissions. Ruminal protein metabolism warrants further research. Harvesting in the wild decreases nutrient loading of marine environment, but effective cultivation and harvesting may impair the equilibrium of coastal ecosystems. ⁷
	Duckweed	Yes	Yes	Yes	30-40	10-18	No	High Can recover nutrients from wastewaters.
970	¹ TInt = Intensive temperate production, TExt = Extensive temperate production, Yes = suitable, (Yes) = suitable with some restrictions such as							
971	species or cultivars (pulses, grass and wheat) or the proximity of the seaside (seaweed)							
972	² Van Krimpen <i>et al.</i> , 2013							
973	³ Bayat <i>et al.</i> , 2015							
974	⁴ Watson <i>et al.</i> , 2017							
975	⁵ Table 3							
976	⁶ Yield potential in tropical areas; Heuzé <i>et al.</i> , 2016b							
977	⁷ Makkar <i>et al.</i> , 2016							

978 **Table 3** *The effect of some alternative protein feeds on milk production of ruminants*

Species	Alternative protein feed ¹	Control protein feed ²	SR ³ %	Diet DMI ⁴ % ⁵	Milk yield in control kg/d	Yield % ⁵				Milk urea % ⁵	N ⁶	Ref. ⁷
						Milk	Lactose	Fat	Protein			
Cow	Camelina E	RSM	100	-3	31	4	4	-3	1	-16	1	1
	Faba bean	RSM	50	-3	31-32	-2	-2	0	-4	7	2	2
	Faba bean	RSM	100	-4	25-35	-6	-5	-2	-7	13	5	2-5
	Faba bean	SBM	40	-1 ⁸	20-22	0	1	-3	-1	-10	2	6
	Faba bean	SBM	100	-1	27	0	1	-3	-1	-10	1	7
	Lupin, blue	RSM	50	-1	31	-4	-3	0	-2	-5	1	8
	Lupin, blue	RSM	100	-4	31-35	-6	-3	2	-6	2	2	4,8
	Lupin, white	SBM	100	-1	26-38	0	1	-1	-3	3	5	9-11
	Lupin, yellow	SBM	100	-5	32	-6	-5	0	-9	nr ⁹	1	12
	Pea	RSM	50	-1	24	-2	-2	1	-3	2	1	13
	Pea	RSM	100	-3	24-25	-6	-6	-5	-7	12	2	5,13
	Pea	RSM-SBM	95	nr	32	-5	nr	6	-2	nr	1	14
	Pea	SBM	33-80	4	21-35	2	3	3	4	17	5	15-17
	Pea	SBM	100	2	21-27	2	3	1	3	-2	2	15,18
	Microalgae	RSM	50	-1	23-31	0	-1	-1	2	4	3	3,19
	Microalgae	RSM	100	0	23-28	-3	-2	-2	-1	3	2	19
	Microalgae	SBM	100	0	30	4	4	11	4	-8	3	20
Sheep	Camelina E	RSM	50-60	nr	1.2	11	-1	-6	-2	nr	2	21,23
	Camelina E	RSM	100	nr	1.2	8	-1	-14	-1	nr	1	21
	Camelina S	SBM	50	-2	0.7-0.8	7	8	11	6	nr	2	23
	Faba bean	SBM	100	2	0.7-0.8	-1	2	-1	2	nr	2	24,25
	Lupin, white	SBM	100	-5	1.4	5	8	3	1	-2	1	26
	Pea	SBM	100	-2	0.7-0.8	9	12	7	4	-2	2	24,25
	Pea	SBS-SFM	100	-5	1.0	4	3	6	8	nr	1	27
Goat	Faba bean	CS	100	0	1.1	-2	-11	-11	0	nr	1	28
	Faba bean	WLS	100	3	1.6	1	-2	-3	0	nr	1	29

979 ¹E = expeller, S = seed

980 ²CON = concentrate mixture, CS cottonseeds, RSM = rapeseed meal, SBM = soybean meal, SBS =
981 soybean seeds, SFM = sunflowerseed meal, WLS = white lupin seeds

982 ³Isonitrogenous substitution rate of control protein feed by alternative protein feed

983 ⁴DMI = dry matter intake

984 ⁵Change (%) due to alternative protein feed compared to control protein feed

985 ⁶Number of diet comparisons

986 ⁷References shown in Supplemental Table S2

987 ⁸Concentrate intake

988 ⁹Not reported

989 **Table 4** *The effect of some alternative feeds on the average daily gains of ruminants*

Species	Alternative feed	Control feed	SR ¹ %	Diet DMI ²	ADG ²	Main findings	Ref. ³
Beef steers	Camelina meal	Soybean meal	100	dec	-	Camelina increased plasma 18:3n-3 concentration and lessened the acute-phase protein reaction.	1
Dairy heifers	Camelina meal	Linseed meal	100	-	-	Camelina decreased plasma insulin concentration.	2
		Distillers dried grains with solubles	100	-	-	Camelina had no major effect on CP or NDF total tract digestibility or rumen fermentation except for higher ammonia relative to other treatments.	
Sheep	Camelina expeller	Rapeseed meal	50 100	nr	nr	Camelina increased muscle t11 18:1, c9t11 18:2 and n-3 fatty acid content, but had no effect on 18:0 or c9 18:1	3
Beef bulls	Lupin (blue) seeds	Rapeseed meal	100	dec	dec	Carcass weight and dressing percentage were the highest for rapeseed. Protein source had no effect on carcass classification or gross chemical composition. Muscle fatty acid profile was similar for lupin and soybean diets, but on rapeseed diet muscle c9t11 18:2 and 18:3n-3 contents were higher.	4
		Soybean meal	100	-	-		
Beef bulls	Lupin (white) seeds	Soybean seeds and meal	100	-	-	Main slaughtering and sectioning characteristics were equal. Lupin diet reduced fatness. Quality traits of meats were comparable in terms of colour, tenderness and chemical and fatty acid profile as well as post slaughtering pH.	5
Beef bulls	Faba bean-cereal silage	Grass silage	100	-	-	Replacing grass silage with grain legume-cereal whole crop silages had no remarkable effect on carcass characteristics, meat quality, fatty acid profile or sensory score.	6
	Pea-cereal silage		100	-	-		
Beef steers	Lupin (white) silage	Grass silage	100	-	-	Carcass merits were equal. Lupin nitrogen degraded faster in the rumen compared to grass.	7

Sheep	Faba beans Lupin (white) seeds	Soybean expeller 100 100	dec dec	- -	Protein source had no effect on carcass characteristics except for decreased back fat thickness for faba bean.	8
Sheep	Lupin (white) seeds	Rapeseed meal Soybean meal 100 100	- -	- -	Digestibility of CP and energy were higher for lupin than rapeseed and soybean.	9
Sheep	Peas	Soybean meal 45 100	- -	- -	Carcass and meat composition and quality were not affected by treatments.	10
Sheep	Pea silage	Grass silage 50	-	inc	Lambs offered pea silage low in tannins grew faster, had increased chop length and improved digestibility of OM and N compared to grass silage as sole forage in the diet.	11
Sheep	Seaweed	Soybean-barley concentrate 20	-	-	Replacing 20 % of soybean-barley concentrate with seaweeds (<i>Ruppia maritima</i> or <i>Chaetomorpha linum</i>) had no effect on OM or CP digestibility, nitrogen partitioning or water intake.	12
Sheep	Seaweed	Alfalfa hay 8 13	- -	- -	Dietary supplementation of seaweed (<i>Ulva lactuca</i>) at low level has no adverse effect on growth of sheep.	13
Goat	Jatropha kernel expeller	Soybean expeller 50 100	inc -	inc -	Replacing 50% or 100% of soybean expeller with fungally detoxified jatropha kernel expeller had no adverse effects on blood parameters. Diet with 1:1 (w/w) soybean expeller and jatropha kernel expeller resulted in highest DM and CP intake and ADG.	14
Sheep	Jatropha expeller	Soybean meal 70	-	-	Replacing 70% of soybean meal in concentrate mixture had no adverse effects on DM intake or ADG of male lambs. The fertility of rams was slightly improved by jatropha inclusion in the diet.	15

990 ¹Substitution rate of control feed by alternative feed

991 ²Effect of alternative feed on dry matter intake (DMI) or average daily gain (ADG): Dec = decrease, - = no effect, inc = increase, nr = not reported

992 ³References shown in Supplemental Table S3

Table 5 *Effect of using tropical fodder tree and shrubs supplementation on feed intake, rumen volatile fatty acid production and milk yield in ruminants fed rice straw based diets.*

Supplement	Form	Dose kg/d	Species	DM ¹ intake	Rumen fermentation ²				Milk yield	Ref. ³
					TVFA	C ₂	C ₃	C ₄		
Cassava	Hay	2.0	Dairy cow	inc ⁴	inc	dec	inc	dec	inc	1
	Silage	2.5	Dairy cow	inc	inc	dec	inc	-	inc	2
Leucaena	Silage	RLS60 ⁵	Dairy steer	inc	inc	dec	inc	-		3
	Hay	6.0	Buffaloes	-	inc	dec	inc	-		4
Flemingia	Hay	FHM+CH ⁶	Dairy steer	-	-	dec	inc	dec		5

¹DM = dry matter

²TVFA = total volatile fatty acids, C₂ = acetate, C₃ = propionate, C₄ = butyrate

³References shown in Supplemental Table S5

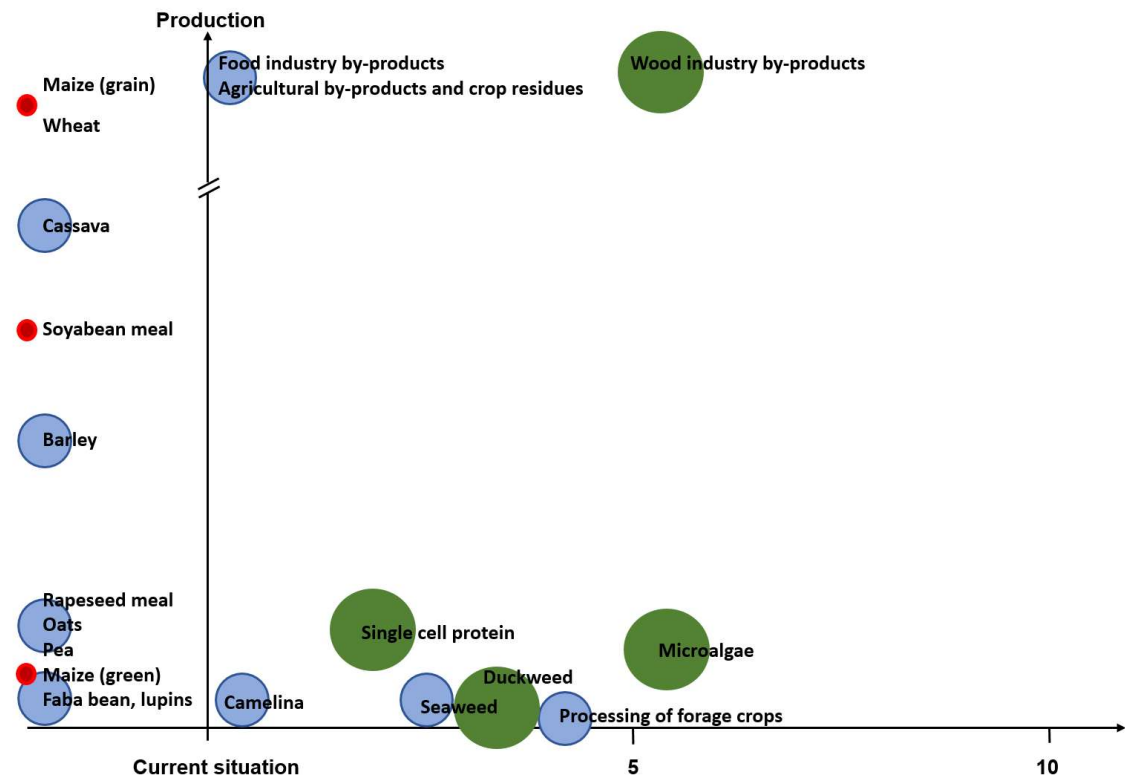
⁴dec = decrease, - = no effect, inc = increase

⁵RLS60 = 40% rice straw + 60% leucaena silage fed ad libitum

⁶FHM+CH = 75 g flemingia hay meal + 75 g cassava hay

List of figure captions

Figure 1 Rough overview of some feeds for ruminants with respect to time to enter readily on the market, extent of production today and potential to increase utilization in ruminant nutrition sustainably in future (small red bubble = limited; medium-sized blue bubble = moderate; large green bubble = high). Data adapted in part from FAOSTAT (2016), Kruus and Hakala (2016) and USDA (2016).



Supplementary File – for Online Publication Only

Review: Alternative and novel feeds for ruminants - nutritive value, product quality and environmental aspects

A. Halmemies-Beauchet-Filleau, M. Rinne, M. Lamminen, C. Mapato, T. Ampapon, M. Wanapat and A. Vanhatalo

Supplemental Table S1 Full references for the chemical composition of some alternative and common feeds for ruminants in Table 1

Feed	References
<i>Common protein feeds</i>	
Rapeseed expeller	Heuzé V, Tran G, Sauvant D, Lessire M and Lebas F 2017. Rapeseed meal. Retrieved on 27 April 2018, from http://www.feedipedia.org/node/52
Soybean expeller	Heuzé V, Tran G and Kaushik S 2017. Soybean meal. Retrieved on 27 April 2018, from http://www.feedipedia.org/node/674
<i>By-products of food industry</i>	
Apple pomace	Wadhwa M, Bakshi MP and Makkar HP 2015. Waste to worth: fruit wastes and by-products as animal feed. CAB Reviews 10, 1-26.
Camelinaseed expeller	Heuzé V, Tran G and Lebas F 2017. Camelina (Camelina sativa) seeds and oil meal. Retrieved on 27 April 2018, from http://www.feedipedia.org/node/4254 Lawrence RD, Anderson JL and Clapper JA 2016. Evaluation of camelina meal as a feedstuff for growing dairy heifers. Journal of Dairy Science 99, 6215-6228.
Cauliflower leaf	Rinne M, Dragomir C, Kuoppala K, Smith J and Yáñez-Ruiz D 2014. Novel feeds for organic dairy chains. Organic Agriculture 4, 275-284.
Cucumber waste	
Grape marc	
Tomato fruit waste	
Olivesilage (pulp + leaf)	
<i>Grain legume seeds</i>	
Faba bean	Heuzé V, Tran G, Delagarde R, Lessire M and Lebas F 2016. Faba bean (Vicia faba). Retrieved on 27 April 2018, from http://www.feedipedia.org/node/4926
Lupins	Berk A, Bamm A, Böhm H, Aulrich K and Rühl G 2008. The nutritive value of lupins in sole cropping systems and mixed intercropping with spring cereals for grain production. In Proceedings of the 12th International Lupin Conference, Lupins for Health and Wealth, 14-18 September 2008, Fremantle, Western Australia, pp. 66-70. Aulrich K and Rühl G 2008. The nutritive value of lupins in sole cropping systems and mixed intercropping with spring cereals for grain production. In Proceedings of the 12th International Lupin Conference, Lupins for Health and Wealth, 14-18 September 2008, Fremantle, Western Australia, pp. 66-70.

	Wasilewko J and Buraczewska L 1999. Chemical composition including content of amino acids, minerals and alkaloids in seeds of three lupin species cultivated in Poland. <i>Journal of Animal and Feed Sciences</i> 81, 1-12.
Pea	Heuzé V, Tran G, Giger-Reverdin S, Noblet J, Renaudeau D, Lessire M and Lebas F 2017. Pea seeds. Retrieved on 27 April 2018, from http://www.feedipedia.org/node/264 http://www.feedipedia.org/node/264
Soybean	Heuzé V, Tran G and Kaushik S 2017. Soybean meal. Retrieved on 27 April 2018, from http://www.feedipedia.org/node/674
<i>Grass silage juice</i>	Franco M, Winqvist E, Rinne M. 2018. Grass silage for biorefinery – A meta-analysis of liquid-solid separation. XVIII International Silage Conference, 24-26 July 2018, Bonn, Germany.
<i>Grain legume whole crop stands</i>	Rinne M, Dragomir C, Kuoppala K, Smith J and Yáñez-Ruiz D 2014. Novel feeds for organic dairy chains. <i>Organic Agriculture</i> 4, 275-284.
<i>Trees or shrubs (leaves unless otherwise stated)</i>	
Cassava Flemingia	Phesatcha B Wanapat M Phesatcha K Ampapon T and Kang S 2016. Supplementation of <i>Flemingia macrophylla</i> and cassava foliage as a rumen enhancer on fermentation efficiency and estimated methane production in dairy steers. <i>Tropical Animal Health and Production</i> 48, 1449-1454.
Leucaena	Phesatcha K and Wanapat M 2017. Tropical legume supplementation influences microbial protein synthesis and rumen ecology. <i>Journal of Animal Physiology and Animal Nutrition</i> 101, 552–562. ‘
Moringa	Makkar HPS and Becker K 1996. Nutritional value and antinutritional components of whole and ethanol extracted <i>Moringa oleifera</i> leaves. <i>Animal Feed Science and Technology</i> 63, 211–228.
Pine bark	Kairenius P, Mäntysaari P and Rinne M 2017. The effect of gradual dietary bark meal supplementation on feed intake and milk production of Nordic Red cows fed a grass silage-based diet. Manuscript.
Sesbania	Teklea D, Gebrua G, Hagosa H and Belay S 2016. Effect of on farm supplementation of dried <i>Sesbania sesban</i> (L.) leaf on performance of Abergelle rams. <i>Scientific Journal of Animal Science</i> 5, 322-328.
Willow	Smith J, Kuoppala K, Yáñez-Ruiz D, Leach K and Rinne M 2014. Nutritional and fermentation quality of ensiled willow from an integrated feed and bioenergy agroforestry system in UK. . In <i>Proceedings of Maataloustieteen Päivät 2014</i> , 8-9 January 2014, Helsinki, Finland. 9 p. Retrieved on 15 December 2017, from http://www.smts.fi/MTP_julkaisu_2014/Posterit/064Smith_ym_Nutritional_and_fermentation_quality_of_ensiled_willow.pdf

<i>Jatropha</i> kernel meal detoxified	Heuzé V, Tran G, Edouard N, Renaudeau D, Bastianelli D and Lebas F 2016. <i>Jatropha</i> (<i>Jatropha</i> sp.) kernel meal and other <i>jatropha</i> products. Retrieved on 30 November 2017, from https://www.feedipedia.org/node/620 https://www.feedipedia.org/node/620
Single-cell protein	
Bacteria	Lindberg JE, Lindberg G, Teräs J, Poulsen G, Solberg SØ, Tybirk K, Przedzymirska J, Sapota GP, Olsen ML, Karlson H, Jóhannsson R, Smáráson BÖ, Gylling M, Knudsen MT, Dorca-Preda T, Hermansen JE, Kruklite Z and Berzina I 2016. Nordic Alternative Protein Potentials: Mapping of regional bioeconomy opportunities. Nordic Council of Ministers. Retrieved on 27 April 2018, from http://www.nordic-ilibrary.org/environment/nordic-alternative-protein-potentials tn2016-527, from http://www.nordic-ilibrary.org/environment/nordic-alternative-protein-potentials tn2016-527
Fungi	
Yeast	Nasseri AT, Rasoul-Amini S, Morowvat MH and Ghasemi Y 2011. Single cell protein: production and process. American Journal of Food Technology 6, 103-116. Ghasemi Y 2011. Single cell protein: production and process. American Journal of Food Technology 6, 103-116.
Microalgae	
<i>Chlorella vulgaris</i>	Lamminen M, Halmemies-Beauchet-Filleau A, Kokkonen T, Simpura I, Jaakkola S, Vanhatalo A 2017. Comparison of microalgae and rapeseed meal as supplementary protein in the grass silage based nutrition of dairy cows. Animal Feed Science and Technology 234, 295-311.
<i>Spirulina platensis</i>	
<i>Euglena gracilis</i>	Aemiro A, Watanabe S, Suzuki K, Hanada M, Umetsu K and Nishida T 2016. Effects of <i>Euglena</i> (<i>Euglena gracilis</i>) supplemented to diet (forage: concentrate ratios of 60: 40) on the basic ruminal fermentation and methane emissions in in vitro condition. Animal Feed Science and Technology 212, 129-135.
<i>Scenedesmus obliquus</i>	Klostermeyer H, Schmandke H, Soeder CJ, Schreiber W, Oehlenschläger J, Scholtyssek S, Kobald M, Sander A, Eilers E, Kries E 2017. Proteins. In Ullmann's Food and Feed (ed. B Elvers), Wiley-VHC, Weinheim, Germany, vol. 2. pp. 861-914., vol. 2. pp. 861-914.
<i>Schizochytrium</i> sp.	Madeira MS, Cardoso C, Lopes PA, Coelho D, Afonso C, Bandarra NM and Prates JA 2017. Microalgae as feed ingredients for livestock production and meat quality: a review. Livestock Science 205, 111-121.
Seaweeds	Makkar HP, Tran G, Heuzé V, Giger-Reverdin S, Lessire M, Lebas F and Ankers P 2016. Seaweeds for livestock diets: a review. Animal Feed Science and Technology 212, 1-17.
Duckweed	Heuzé V and Tran G 2015. Duckweed. Retrieved on 26 July 2017, from https://www.feedipedia.org/node/15306

12 **Supplemental Table S2** *Full references for Table 3 reporting the effect of some alternative*
 13 *protein feeds on the milk production of ruminants*

No.	Full reference
1	Halmemies-Beauchet-Filleau A, Kokkonen T, Lampi AM, Toivonen V, Shingfield KJ and Vanhatalo A 2011. Effect of plant oils and camelina expeller on milk fatty acid composition in lactating cows fed diets based on red clover silage. <i>Journal of Dairy Science</i> 94, 4413–4430.
2	Puhakka L, Jaakkola S, Simpura I, Kokkonen T and Vanhatalo A 2016. Effects of replacing rapeseed meal with fava bean at 2 concentrate crude protein levels on feed intake, nutrient digestion, and milk production in cows fed grass silage-based diets. <i>Journal of Dairy Science</i> 99, 7993–8006.
3	Halmemies-Beauchet-Filleau A, Lamminen M, Kokkonen T, Vanhatalo A and Jaakkola S 2016. Rapeseed meal, faba beans and microalga (<i>Spirulina platensis</i>) as protein supplements for dairy cows on grass silage based diets. In <i>Proceedings of 5th EAAP International Symposium on Energy and Protein Metabolism and Nutrition</i> , 12-15 September 2016, Krakow, Poland pp. 281-283.
4	Kuoppala K, Jaakkola S, Ahvenjärvi S and Rinne M 2016. Härkäpapu ja sinilupiini lypsylehmien valkuaisrehuna. In <i>Proceedings of Maataloustieteen Päivät 2016</i> , 12-13 January 2016, Helsinki, Finland. Retrieved on 15 December 2017, from p. 27. http://www.smts.fi/sites/smts.fi/files/MAATALOUSTIETEEN_ABSTRAKTIKIRJA2016.pdf
5	Ramin M, Höjer A and Hetta M 2017. The effects of legume seeds on the lactation performance of dairy cows fed grass silage-based diets. <i>Agricultural and Food Science</i> 26, 129-137.
6	Volpelli LA, Comellini M, Masoero F, Moschini M, Lo Fiego DP and Scipioni R 2010. Faba beans (<i>Vicia faba</i>) in dairy cow diet: effect on milk production and quality. <i>Italian Journal of Animal Science</i> , 9, e27.
7	Tufarelli V, Khan RU and Laudadio V 2012. Evaluating the suitability of field beans as a substitute for soybean meal in early-lactating dairy cow: Production and metabolic responses. <i>Animal Science Journal</i> 83, 136-140.
8	Partially published in Puhakka L, Jaakkola S, Kokkonen T and Vanhatalo A 2017. Blue lupin as an alternative protein supplement for dairy cows fed grass silage-based diets. In <i>Proceedings of NJF Seminar 495</i> , 19-21 June 2017, Mikkeli, Finland pp. 80.
9	Singh CK, Robinson PH and McNiven MA 1995. Evaluation of raw and roasted lupin seeds as protein supplements for lactating cows. <i>Animal Feed Science and Technology</i> 52, 63-76.
10	Robinson PH and McNiven MA 1993. Nutritive value of raw and roasted sweet white lupins (<i>Lupinus albus</i>) for lactating dairy cows. <i>Animal Feed Science and Technology</i> 43, 275-290.
11	Froidmont E and Bartiaux-Thill N 2004. Suitability of lupin and pea seeds as a substitute for soybean meal in high-producing dairy cow feed. <i>Animal Research</i> 53, 475-487.
12	Marley C, Davies D, Fisher B, Fychan R, Sanderson R, Jones R and Abberton M 2008. Effects of incorporating yellow lupins into concentrate diets compared with soya on milk production and milk composition when offered to dairy cows. In <i>Proceedings of the 12th International Lupin Conference—Lupins for health and wealth</i> , 14-18 September 2008, Fremantle, Western Australia pp. 115-117.
13	Khalili H, Kuusela E, Suvitie M and Huhtanen P 2002. Effect of protein and energy supplements on milk production in organic farming. <i>Animal Feed Science and Technology</i> 98, 103-119.
14	Corbett RR, Goonewardene LA and Okine EK 1995. Effects of feeding peas to high-producing dairy cows. <i>Canadian Journal of Animal Science</i> 75, 625-629.
15	Khorasani GR, Okine EK, Corbett RR, Kennelly JJ 2001. Nutritive value of peas for lactating dairy cattle. <i>Canadian Journal of Animal Science</i> 81, 541–551.
16	Petit HV, Rioux R and Ouellet DR 1997. Milk production and intake of lactating cows fed raw or extruded peas. <i>Journal of Dairy Science</i> 80, 3377-3385.

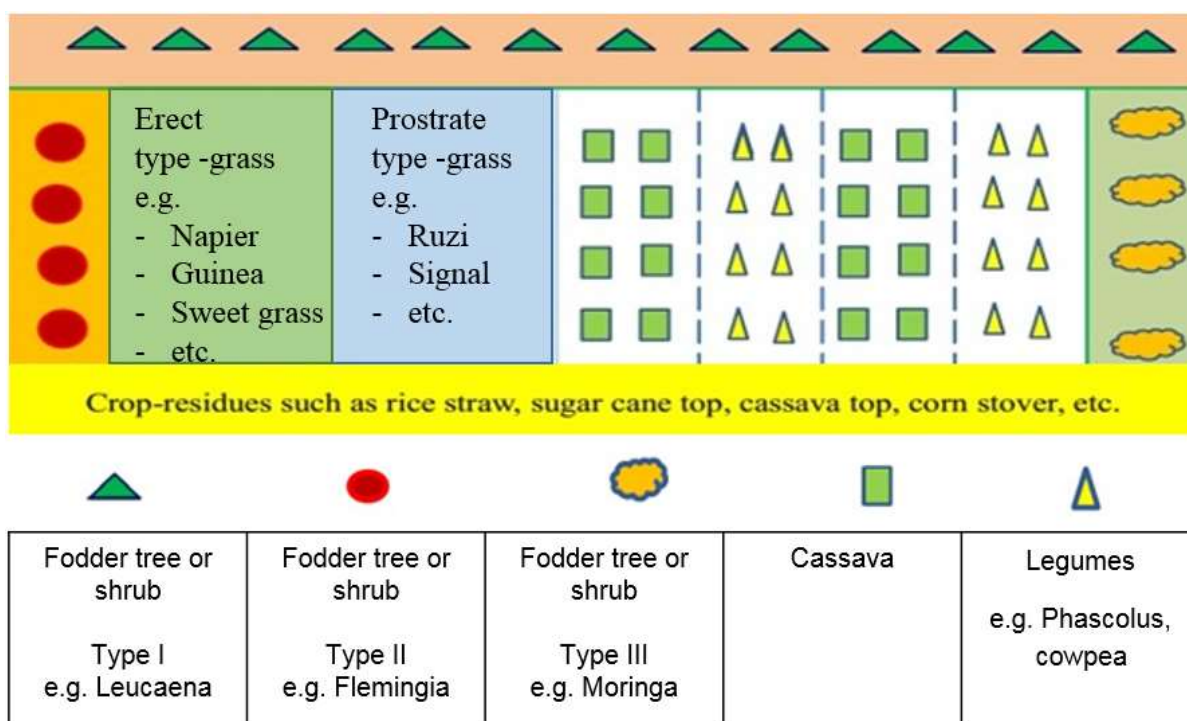
- 17 Vander Pol M, Hristov AN, Zaman S and Delano N 2007. Peas can replace soybean meal and corn grain in dairy cow diets. *Journal of Dairy Science* 91, 698-703.
- 18 Tufarelli V, Naz S, Khan RU, Mazzei D and Laudadio V 2012. Milk quality, manufacturing properties and blood biochemical profile from dairy cows fed peas (*Pisum sativum* L.) as dietary protein supplement. *Tierzucht* 55, 132-139.
- 19 Lamminen M, Halmemies-Beauchet-Filleau A, Kokkonen T, Simpura I, Jaakkola S and Vanhatalo A 2017. Comparison of microalgae and rapeseed meal as supplementary protein in the grass silage based nutrition of dairy cows. *Animal Feed Science and Technology* 234, 295-311.
- 20 Lamminen M, Halmemies-Beauchet-Filleau A, Kokkonen T, Jaakkola S and Vanhatalo A 2016. Microalgae as a substitute for soya bean meal in the grass silage based dairy cow diets. In *Proceedings of 5th EAAP International Symposium on Energy and Protein Metabolism and Nutrition*, 12-15 September 2016, Krakow, Poland pp. 285-287.
- 21 Szumacher-Strabel M, Cieślak A, Zmora P, Pers-Kamczyc E, Bielińska S, Stanisław M and Wójtowski J 2011. Camelina sativa cake improved unsaturated fatty acids in ewe's milk. *Journal of the Science of Food and Agriculture* 91, 2031-2037.
- 22 Danków R, Pikul J, Wójtowski J, Cais-Sokolińska D, Teichert J, Bagnicka E, Cieslak A and Szumacher-Strabel M 2015. The effect of supplementation with gold of pleasure (*Camelina sativa*) cake on the fatty acid profile of ewe milk and yoghurt produced from it. *Journal of Animal and Feed Sciences* 24, 193-202.
- 23 Mierlita D and Vicas S 2015. Dietary effect of silage type and combination with camelina seed on milk fatty acid profile and antioxidant capacity of sheep milk. *South African Journal of Animal Science* 45, 1-11.
- 24 Liponi GB, Casini L, Martini M and Gatta 2007. Faba bean (*Vicia faba minor*) and pea seeds (*Pisum sativum*) as protein sources in lactating ewes' diets. *Italian Journal of Animal Science* 6(sup1), 309-311.
- 25 Bonanno A, Di Grigoli A, Vitale F, Alabiso M, Giosuè C, Mazza F and Todaro M 2016. Legume grain-based supplements in dairy sheep diet: effects on milk yield, composition and fatty acid profile. *Animal Production Science* 56, 130-140.
- 26 Masucci F, Di Francia A, Romano R, di Serracapriola MM, Lambiase G, Varricchio ML and Proto V 2006. Effect of *Lupinus albus* as protein supplement on yield, constituents, clotting properties and fatty acid composition in ewes' milk. *Small Ruminant Research* 65, 251-259.
- 27 Renna M, Cornale P, Lussiana C, Malfatto V, Fortina R, Mimosi A and Battaglini LM 2012. Use of *Pisum sativum* (L.) as alternative protein resource in diets for dairy sheep: effects on milk yield, gross composition and fatty acid profile. *Small Ruminant Research* 102, 142-150.
- 28 Sampelayo MS, Pérez ML, Extremera FG, Boza JJ and Boza J 1999. Use of Different Dietary Protein Sources for Lactating Goats: Milk Production and Composition as Functions of Protein Degradability and Amino Acid Composition¹. *Journal of Dairy Science* 82, 555-565.
- 29 Morales ER, Alcaide EM and Sampelayo MR 2008. Milk production of dairy goats fed diets with different legume seeds: effects of amino acid composition of the rumen undegradable protein fraction. *Journal of the Science of Food and Agriculture* 88, 2340-2349.

15 **Supplemental Table S3** Full reference for Table 4 reporting the effect of some alternative feeds on the average daily gains of ruminants

No.	Full reference
1	Cappellozza BI, Cooke RF, Bohnert DW, Cherian G and Carroll JA 2012. Effects of camelina meal supplementation on ruminal forage degradability, performance, and physiological responses of beef cattle. <i>Journal of Animal Science</i> 90, 4042-4054.
2	Lawrence RD, Anderson JL and Clapper JA 2016. Evaluation of camelina meal as a feedstuff for growing dairy heifers. <i>Journal of Dairy Science</i> 99, 6215-6228.
3	Cieslak A, Stanisz M, Wojtowski J, Pers-Kamczyc E, Szczechowiak J, El-Sherbiny M and Szumacher-Strabel M 2013. Camelina sativa affects the fatty acid contents in M. longissimus muscle of lambs. <i>European Journal of Lipid Science and Technology</i> 115, 1258-1265.
4	Sami AS, Schuster M and Schwarz FJ 2009. Performance, carcass characteristics and chemical composition of beef affected by lupine seed, rapeseed meal and soybean meal. <i>Journal of Animal Physiology and Animal Nutrition</i> 94, 465-473.
5	Vicenti A, Totada F, Di Turi L, Cocca C, Perrucci M, Melodia L and Ragni M 2009. Use of sweet lupin (<i>Lupinus albus</i> L. var. Multitalia) in feeding for Podolian young bulls and influence on productive performances and meat quality traits. <i>Meat Science</i> 82, 247-251.
6	Huuskonen A, Pesonen M and Honkavaara M 2016. Performance and meat quality of Nordic Red and Aberdeen Angus bulls offered faba bean or field pea based whole crop legume-cereal silages. <i>Agricultural and Food Science</i> 25, 1-12.
7	Murphy SR, McNiven MA, MacLeod JA and Halliday LJ 1993. Grass and lupin silage in rations for beef steers supplemented with barley or potatoes. <i>Animal Feed Science and Technology</i> 40, 273-283.
8	Purroy A, Echaide H, Muñoz F, Arana A and Mendizabal JA 1993. The effect of protein level and source of legume seeds on the growth and fattening of lambs. <i>Livestock Production Science</i> 34, 93-100.
9	Stanford K, Lees BM, McAllister TA, Xu ZJ and Cheng KJ 1996. Comparison of sweet white lupin seed, canola meal and soybean meal as protein supplements for lambs. <i>Canadian Journal of Animal Science</i> 76, 215-219.
10	Lanza M, Bella M, Priolo A and Fasone V 2003. Peas (<i>Pisum sativum</i> L.) as an alternative protein source in lamb diets: growth performances, and carcass and meat quality. <i>Small Ruminant Research</i> 47, 63-68.
11	Hart KJ, Sinclair LA, Wilkinson RG and Huntington JA 2011. Effect of whole-crop pea (L.) silages differing in condensed tannin content as a substitute for grass silage and soybean meal on the performance, metabolism, and carcass characteristics of lambs. <i>Journal of Animal Science</i> 89, 3663-3676.
12	Ktita SR, Chermiti A and Mahouachi M 2010. The use of seaweeds (<i>Ruppia maritima</i> and <i>Chaetomorpha linum</i>) for lamb fattening during drought periods. <i>Small Ruminant Research</i> 91, 116-119.
13	El-Waziry A, Al-Haidary A, Okab A, Samara E and Abdoun K 2015. Effect of dietary seaweed (<i>Ulva lactuca</i>) supplementation on growth performance of sheep and on in vitro gas production kinetics. <i>Turkish Journal of Veterinary and Animal Sciences</i> 39, 81-86.
14	Belewu MA, Belewu KY and Lawal IA 2013. Cocktail of fungi blend on <i>Jatropha curcas</i> kernel cake: effect on feed intake and blood parameters of goat. <i>American-Eurasian Journal of Agricultural and Environmental Sciences</i> 13, 315-320.
15	El-Zelaky OA, Khalifa EI, Mohamed AH, Bahera KM and Hussein AM 2011. Productive and reproductive performance of rahmani male lambs fed rations containing jatropha cake. <i>Egyptian Journal of Sheep and Goat Sciences</i> 6, 15-24.

Supplemental Table S4 *Full reference for Table 5 reporting the effect of using tropical fodder tree and shrubs supplementation on animal performance*

No.	Full reference
1	Wanapat M and Kang S 2013. Enriching the nutritive value of cassava as feed to increase ruminant productivity. <i>Journal of Nutritional Ecology and Food Research</i> 1, 262-269.
2	Wanapat M, Phesatcha K, Viennasay B, Kang S 2016. Performance of tropical dairy cows fed on cassava top silage in rice straw based diet. In <i>Proceedings of the 17th AAAP Animal Science Congress</i> , 22-25 August 2016, Fukuoka, Japan, pp. 201-206.
3	Giang N, Truong T, Wanapat M, Phesatcha K and Kang S 2016. Level of <i>Leucaena leucocephala</i> silage feeding on intake, rumen fermentation, and nutrient digestibility in dairy steers. <i>Tropical Animal Health and Production</i> 48, 1057-1064.
4	Phesatcha K and Wanapat M 2016. Tropical legume supplementation influences microbial protein synthesis and rumen ecology. <i>Journal of Animal Physiology and Animal Nutrition</i> 101, 552–562.
5	Phesatcha B, Wanapat M, Phesatcha K, Ampapon T and Kang S 2016. Supplementation of <i>Flemingia macrophylla</i> and cassava foliage as a rumen enhancer on fermentation efficiency and estimated methane production in dairy steers. <i>Tropical Animal Health and Production</i> 48, 1449-1454.



Supplemental Figure S1 Proposed sustainable ruminant feeding system for smallholder farmers in the tropics

Reference: Wanapat M, Foiklang S, Ampapon T, Mapato C and Cherdthong T 2017. Feeding strategy on farms to improve livestock productivity and reduce methane production. In Proceedings of the 2nd International Conference on Animal Nutrition and Environment, 1-4 November 2017, Khon Kaen, Thailand, pp. 14-29.